



REVIEW

Brain Mechanisms Underlying Panic Attack and Panic Disorder

Xuyan Guan^{1,2} · Peng Cao^{1,2}

Received: 6 February 2023 / Accepted: 23 April 2023 / Published online: 21 July 2023

© Center for Excellence in Brain Science and Intelligence Technology, Chinese Academy of Sciences 2023

Abstract Panic disorder is a psychiatric disorder characterized by recurrent panic attacks, with a prevalence of ~4% in the general population, causing heavy personal and socioeconomic burdens. The similarities of animal defense responses to clinical panic attack symptoms in humans make it possible to translate neuroanatomical pathways identified in animal studies to panic disorder in humans. Therefore, in this review we first present a basic overview of panic disorder in humans including the main subtypes, models commonly used to trigger panic attacks, related hypotheses, the neurotransmitter systems that may be involved, and the current clinical treatments to give the reader a comprehensive understanding of panic disorder. The animal section introduces the models that trigger panic-like behavior in animals and the brain regions that may be involved, providing insights for future elucidation of the neural circuit mechanisms behind panic attacks.

Keywords Panic disorder · Panic attack · Defense responses · Animal studies · Humans

Introduction

With the publication of the Diagnostic and Statistical Manual of Mental Disorders III (DSM) in 1980, panic disorder (PD) was recognized as a diagnostic entity and attracted significant attention [1]. The hallmark of PD is unpredictable

and recurrent panic attacks (PAs). According to the DSM, a PA is defined as a period of intense distress or discomfort characterized by at least four of the following 13 typical symptoms: palpitations, trembling, sweating, choking, chest tightness, dyspnea, nausea, dizziness, paresthesia or numbness, chills or hot flashes, depersonalization/derealization, going crazy, and feeling like one is dying (Fig. 1) [2]. Repeated PAs have a serious negative impact on patients' social and work life. Although basic and clinical research on PD has garnered increasing attention in the field of neuropsychiatry, it remains important to strengthen the diagnosis, treatment, and prevention of PD.

However, studies that can be performed in humans are limited and current PA diagnosis is primarily based on verbal reports, which cannot be obtained from animal studies. These limitations have hindered our understanding of the pathogenesis of PD. Studies have shown that when animals are faced with imminent threats or natural enemies, they display a series of defense responses accompanied by marked autonomic nervous system (ANS) symptoms, such as tachycardia and elevated blood pressure, akin to PAs in humans. Therefore, the study of panic-like behaviors in animals can help to elucidate the pathogenesis of PD.

Concepts of Anxiety, Fear, and Panic

The ability to flexibly choose the optimal defense strategy in response to different threats is critical for survival. Thus, our brain needs to evaluate environmental threats and make appropriate responses to mitigate or avoid harm [3]. During this process, several different emotions i.e., anxiety, fear, and panic are generated accordingly. Various animal studies have shown that different defensive behaviors can be triggered based on 'defensive distance' to the threat and contextual

✉ Xuyan Guan
guanxuyan@nibs.ac.cn

¹ Tsinghua Institute of Multidisciplinary Biomedical Research, Tsinghua University, Beijing 102206, China

² National Institute of Biological Sciences (NIBS), Beijing 102206, China

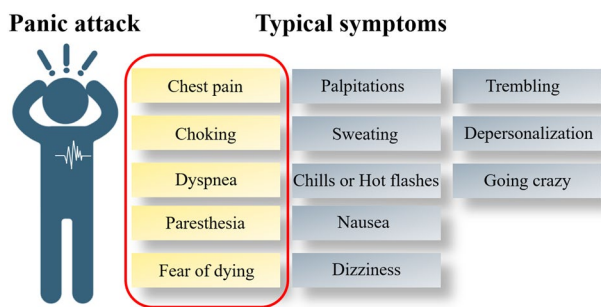


Fig. 1 Typical symptoms of panic attacks. A PA is defined as a period of intense distress or discomfort characterized by at least four of the 13 typical symptoms listed above. In the red box are the main symptoms present in PD patients of the respiratory subtype.

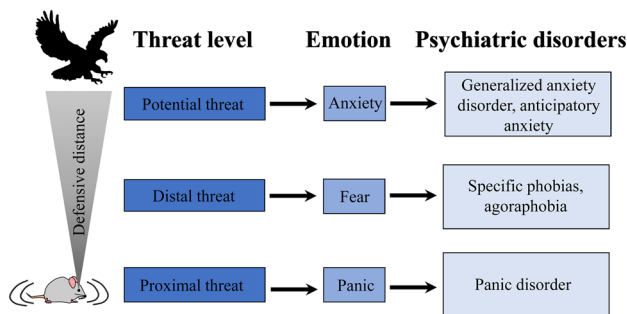


Fig. 2 Overview of the main emotions produced by animals at different threat levels and the associated mental disorders in humans.

conditions, including hiding places and escape routes [4–6]. According to research on wild rats subjected to threatening stimuli [7], there are three main threat levels, i.e., potential, distal, and proximal threats (Fig. 2).

As the first level of defense, a potential threat is defined as when there is no apparent threat or when predators have already been encountered. In this situation, anxiety becomes dominant, and individuals exhibit persistent foraging behavior or adopt risk-assessed defense strategies. It is commonly believed that generalized anxiety disorder (GAD) and anticipatory anxiety are associated with primary defenses, and research has confirmed that antidepressants alleviate these types of psychiatric disorders [8].

Distal threat occurs when the prey has spotted a predator, but the distance between them is large, ensuring relative safety. In this situation, anxiety is replaced by fear and the animal becomes nervous and freezes to reduce the chance of detection by the predator. Animals flee the conditioned aversive stimulus or environment if an exit is available. Fear-related psychiatric disorders, including specific phobias and agoraphobia, for which there are no effective medications, can be cured to some extent by exposure therapy [9].

Proximal threat occurs when the distance between prey and predator is so close that the prey animal has no choice

but to fight or flee. In this situation, the animal becomes extremely panicked, which can be alleviated by the pre-injection of drugs clinically used to treat PD in humans [10]. Indeed, most animal studies do not distinguish between fear and panic.

Brain activity has been found to shift from the forebrain to the midbrain as threat imminence increases [11]. Distal, unpredictable threats activate the prefrontal cortex and cause anxiety [12], while proximal threats activate the periaqueductal gray (PAG), initiating panic-like behavior [4]. From an evolutionary perspective, higher brain processes are responsible for more complex anxiety-related behaviors in response to distal threats, while phylogenetically earlier midbrain regions coordinate rapid and direct avoidance responses. This concept has been validated relatively well in humans. Notably, Mobbs and colleagues developed a new model of active avoidance in humans to study the spatial imminence of threats [13]. In this paradigm, volunteers were chased by a virtual predator (a red circle) within a two-dimensional maze, and once captured received either a low- or high-intensity electrical shock as punishment. Functional magnetic resonance imaging (fMRI) results showed that elevated brain blood flow (reflecting neuronal activity) transferred from the frontal cortex to the cerebellum and PAG, consistent with animal models of defensive avoidance [14, 15]. These results describe a dynamic configuration involving the forebrain to the PAG in response to different levels of threat. Thus, dysfunction in these circuits may contribute to some psychiatric disorders. For example, PAG dissociation from cortical-amygdala regulation may contribute to PD, as manifested by excessive somatic and autonomic fear responses to neutral stimuli [16].

Human Studies

Respiratory and Non-respiratory Panic Subtypes

Currently, PD patients are mainly treated clinically by pharmacological or psychotherapeutic programs with good efficacy, but the optimal intervention for each patient is still uncertain [17], so distinguishing different subtypes of PD according to different clinical symptoms can provide a scientific basis for physicians to develop more individualized treatment plans. A single PA involves several kinds of somatic symptoms, such as respiratory (choking and dyspnea), cardiac (palpitations and chest pain), gastrointestinal (nausea), and vestibular (dizziness) symptoms [2]. Correspondingly, four main subtypes have been reported, namely cardiac, respiratory, gastrointestinal, and vestibular subtypes [18, 19]. Here, we focused mainly on

the respiratory subtype (RS), as it seems to be the most described and studied.

Historically, Klein noted that while respiratory symptoms such as dyspnea are a core feature of PD, this is not true of all PD patients, some showing prominent peripheral autonomic issues such as palpitations and tremors [20]. In a previous study, Briggs and colleagues divided PD patients into two groups based on their description of a recent PA: patients in the RS group typically experienced symptoms such as dyspnea, chest pain or discomfort, smothering sensation, feeling like one is dying, and paresthesia (numbness or tingling sensation) during a single PA (Fig. 1), they also suffered greater work impairment, more spontaneous PAs and fewer situational PAs, as opposed to patients in the non-respiratory subtype (NRS). RS and NRS patients have more effective remission with antidepressants and benzodiazepines, respectively [21].

Many studies have since identified marked differences between RS and NRS [22, 23]. For example, RS patients usually have a later onset than NRS patients and a family history of PD, while NRS patients have significantly more previous episodes of depression [24]. High-resolution MRI studies have shown structural abnormalities of cortical thinning in the superior frontal, caudal-mid frontal, and posterior parietal areas of the left hemisphere of patients with RS, which may be an important pathophysiological mechanism of RS [25]. In addition, PD patients in different subgroups report different symptoms during PA induced by the administration of the same challenge agent. For example, RS patients are more sensitive and responsive to CO₂ than NRS patients [17], and this appears to be the most significant feature that distinguishes RS from NRS.

Although the above evidence suggests that RS and NRS patients are separate subtypes of PD, there is still no unified criterion for the precise definition and treatment of respiratory subtypes of PD, as limitations and variability in methods have led to widely varying results in different studies [26]. More importantly, despite attempts to classify PD into different subtypes, studies have inevitably found that there is still much overlap. For instance, the severity of panic and anxiety can be scored by the Panic Disorder Severity Scale and Beck Anxiety Index, but no differences have been found between the two subtypes [27]. In addition, studies have reported no difference in the therapeutic effects of cognitive behavioral therapy (CBT) [26] and anti-panic drugs such as alprazolam, clonazepam, and nortriptyline on RS and NRS [24, 27, 28]. In the future, standard study methods and the incorporation of multiple determination criteria should be established to distinguish different PD subtypes, which is crucial for achieving precise treatment.

Experimental Models of PA in Humans

Spontaneous PAs are often unprompted and may occur several times a week or only once every few months, making them difficult to study. Therefore, rather than waiting for patients to manifest spontaneous PAs, modeling PAs can be a good solution to this problem. A valid and reliable model should satisfy the following five criteria [29].

(1) Safety: as a prerequisite for any project involving human subjects, symptoms induced by experimental procedures should be transient, easily reversible, and present no potential health risks. (2) Convergence: evoked symptoms should be similar to those of naturally-occurring PAs for subsequent studies to be meaningful. (3) Discrimination: specific distinctions between healthy and pathological subjects should be possible. (4) Reproducibility: model effects should be replicable. (5) Clinical validation: drugs and techniques clinically used to treat PD should alleviate or almost eliminate the induced abnormal symptoms.

Based on these criteria, we focused on four models currently used to trigger PAs in the clinic: i.e., CO₂ exposure, sodium lactate administration, voluntary hyperventilation, and CCK-4 challenge.

CO₂ Exposure

As a metabolite of the human body, CO₂ plays an irreplaceable role in physiological processes. Inhalation of a certain concentration of CO₂ leads to a rapid increase in the partial pressure of intravascular CO₂ (pCO₂) and a decrease in brain pH, which can trigger PAs [30].

Over the past few decades, the inhalation of CO₂ under controlled laboratory conditions has shown numerous advantages over other challenges. Not only is the inhalation of CO₂ a relatively simple and non-invasive procedure, but it is also effective in inducing typical PA symptoms while being of short duration and without adverse health effects in humans [31]. The experimental panicogenic properties of CO₂ can be traced back to 1919. Notably, Drury *et al.* found that patients with “irritable hearts” (similar to PD) are more susceptible to CO₂ than healthy individuals [32]. Similarly, Cohen *et al.* reported that, unlike controls, patients with “neurocirculatory debility” (similar to PD) experienced feelings of fear and anxiety following injection with a 4% CO₂ mixture for 12 min [33]. Many other studies have also confirmed that inhalation of CO₂ at certain concentrations can trigger PA in patients with PD, thereby establishing the CO₂ challenge as a reliable and extensively used model for PD [34].

When it comes to the specific concentration of inhaled CO₂, Bailey *et al.* reported that low doses such as 7.5% CO₂ elicit responses closer to GAD than to PD [35]. In this regard, Perna *et al.* have amply demonstrated that inhalation of 35% CO₂ is a reliable laboratory challenge, highly specific

and stable in patients with PD [36–40]. Using this model, Schruers *et al.* conducted numerous studies. For example, they found that selective 5-hydroxytryptamine reuptake inhibitors (SSRIs) act as anti-panic agents primarily by increasing the availability of serotonin in the brain [41–43]. In addition, 35% CO₂ specifically triggers PAs without activating the hypothalamic-pituitary-adrenal axis, making it a valuable model for studying panic [44]. Therefore, subsequent studies refer to the work of Perna and Schruers to standardize the use of CO₂ inhalation models in humans.

Sodium Lactate Administration

Lactate accumulates in the body during strenuous exercise and subsequently causes muscle soreness, has long been thought of as a waste product for cells to breathe and metabolize in the absence of oxygen. However, it has been found that lactate can be used as an important energy source for neurons [45]. Furthermore, emerging evidence suggests that lactate is also a major recyclable carbohydrate fuel in mammals [46].

Given the abnormal lactate metabolism and exercise intolerance seen in patients with anxiety neurosis under standardized workloads, Pitts *et al.* speculated that lactate itself may be directly related to anxiety attacks [47]. In a double-blind trial using sodium lactate infusion, 13 out of 14 anxious neurotic patients exhibited typical anxiety episodes, compared with only two out of 10 healthy controls [48]. Wiese *et al.* found that sodium lactate infusion was remarkably more likely to trigger PAs in patients with PD than without PD, thus providing a reliable distinction between them [49].

Currently, most studies still follow the standardized sodium lactate injection protocols established by Pitts and McClure [47]. In these experiments, an infusion of 0.5 M sodium lactate (usually in D- or L-lactate form) is initiated at a total dose of 10 mL/kg body weight without patient knowledge until the patient panics. In PD patients, lactate infusion produces classic panic symptoms such as tachycardia and shortness of breath [50, 51]. Although sodium lactate infusion may effectively trigger PAs *via* the same pH-related mechanisms as 35% CO₂ inhalation, it is clear that the latter is more advantageous. Sodium lactate infusion has been largely abandoned due to its need for intravenous lancing and its long duration (up to 40 min) [49].

Voluntary Hyperventilation

The relationship between hyperventilation and PD was intensely debated in the 1980s, with some suggesting that respiratory alkalosis due to hyperventilation is a key cause of PA, while others considered hyperventilation to be an important consequence of a PA [52, 53]. Certainly, there is

a significant link between hyperventilation and PAs, prompting the development of voluntary hyperventilation as a major model for triggering PAs [54].

During the hyperventilation task, participants are generally subjected to rising and falling tones through headphones, with a rise in pitch on inhalation and a fall in pitch on exhalation. This causes a rapid drop in blood pCO₂ levels, resulting in typical sensations such as dizziness, paresthesia, racing heart rate, and breathlessness, which begin as soon as the end-tidal partial pressure of CO₂ (PetCO₂) drops below ~30 mm Hg [55].

Studies have demonstrated that hyperventilation triggers PAs at a prominently higher rate in PD patients than in healthy individuals and patients with other psychiatric disorders, and thus can be used as a simple and rapid diagnostic method [56–59]. Of note, Nardi *et al.* found that RS patients are more sensitive to the breathing challenge test than other PD patients [60–62]. In addition, voluntary hyperventilation has been used for interoceptive exposure to achieve relief by having patients repeatedly experience these associated symptoms (albeit not very efficiently) [54]. However, the panic-like response triggered by voluntary hyperventilation is relatively weak compared to CO₂ inhalation and lactate and is more variable between subjects [63]. Given its ease of use and non-invasive nature, it is often used as a safe and simple test to verify the diagnosis of certain specific patients with PD.

CCK-4 Challenge

Cholecystokinin, an intestinal hormone first discovered and defined in 1928 [64], promotes gallbladder contraction and bile release, thereby facilitating intestinal digestive function [65]. Correspondingly, the CCK-1 receptor subtype is predominantly expressed in the alimentary tract, while the other receptor, CCK-2, is predominantly enriched in the brain [65].

The C-terminal tetrapeptide fragment of CCK (CCK-4), is a naturally-occurring neurotransmitter that has panicogenic effects [52]. In 1968, while studying the effects of gastrin on insulin secretion, Rehfeld unexpectedly experienced full-blown PAs about 30 s after administering CCK-4 intravenously to himself, with symptoms lasting for about 15 min [66]. Inspired by this, Jacques Bradwejn subsequently conducted a large number of systematic studies and showed that intravenous CCK-4 triggers strong PA-related symptoms in both PD patients and controls, the former being more sensitive [67–71]. In terms of pharmacology, imipramine [72], fluvoxamine [73], and citalopram [74] have been shown to alleviate CCK-4-induced PAs. Moreover, positron emission tomography (PET) [75] and fMRI [76, 77] data showed increased activity in the anterior cingulate cortex

(ACC), parts of the cerebral cortex, amygdala, cerebellum, and brainstem after CCK-4 injection.

Currently, CCK-4 challenges are usually administered intravenously [66]. It has been recognized as a reliable and safe model for studying PD in humans and for screening novel anxiolytic drugs [78].

Other Experimental Challenges

In addition to the three approaches described above, several other techniques have been developed to trigger spontaneous PAs in PD patients, including doxapram, yohimbine, caffeine, and *m*-chlorophenyl piperazine (m-CPP).

Given the possible involvement of noradrenergic activity in PD [79], studies have shown that yohimbine, an α 2 adrenoreceptor antagonist [80], triggers somatic anxiety symptoms and cardiovascular responses in PD patients [81]. However, the fact that PD may be caused by multiple nervous system dysfunctions rather than by damage to a single neurotransmitter system limits the use of this model. A study comparing the panicogenic effects of m-CPP (a serotonergic agonist) and caffeine (an adenosine antagonist) found that both triggered significant panic symptoms in PD patients under double-blind conditions [82]. In addition, doxapram, a central nervous system (CNS) stimulant (analeptic), is also considered an effective panicogenic agent that can differentiate between PD patients and healthy people [83, 84].

Although the above challenges have the potential to serve as valid models for initiating PAs in humans, their specific mechanisms of action and reliability need to be further investigated.

Main Hypotheses for PD

As PD is a complex psychiatric disorder, not all patients show stable and positive outcomes after first-line treatment [59]. Thus, to improve treatment and administration, it is crucial to identify the key mechanisms underpinning PD. Although this is still poorly understood after decades of research, some hypotheses about the pathophysiology of PD have been proposed based on data from clinical PD treatments and animal research.

An influential theory, the neuroanatomical hypothesis of PD, was proposed by Gorman *et al.* in 1989 and was revised in 2000. This theory posits that PAs result from dysfunction of the brain's "fear network", centered on the amygdala and includes its inputs and outputs, such as the PAG, hypothalamus, and brainstem [66]. Their landmark work sets the framework for subsequent neurobiological studies of PD and anxiety. However, with the accumulation of relevant studies, the neuroanatomical hypothesis needs to be specifically reassessed. In this regard, Dresler *et al.* made a comprehensive summary and analysis of relevant

studies, as well as a review and critical discussion of the neuroanatomical hypothesis of PD [85]. For example, given the rather rare findings of amygdala hyperactivity in PD patients, its central role in the fear network may need to be reassessed. Conversely, based on brain imaging studies, the importance of some other structures such as the insula or ACC, may have been initially downplayed [85].

The "false suffocation alarm theory" states that everyone is equipped with a brain alarm system to indicate a lack of useful air [20]. In PD patients, this alarm system is falsely triggered in situations that are not truly life-threatening, leading to PAs and mainly characterized by dyspnea [20]. High levels of CO₂ provide such a false signal. Subsequently, Preter *et al.* updated the false suffocation alarm theory, hypothesizing that PD may result from a defect in endogenous opioid function, considering that both CO₂ sensitivity and PAs are controlled by the opioid system [86]. However, the hypothesis is currently largely untested. According to the hypothesis, respiratory symptoms are specific to PD patients, distinguishing them from PA in non-PD patients, but inconsistent results have been reported in some studies [87].

In psychology, the learning theory and the cognitive theory are more prominent. Classical conditioning theories refer to the learning of conditioned reflexes that occurs during the constant pairing of conditioned stimuli (CSs such as rapid heartbeat and dyspnea) and unconditioned aversive stimuli (i.e., PA), causing the reappearance of CSs to trigger panic and anxiety [88]. A further modification of the learning theory was made by Bouton *et al.* who suggested that interoceptive conditioning may be key to the development of PD [89].

Based on the superiority of CBT in the treatment of PD, the cognitive theory has been rapidly developed and widely accepted [90]. It has been proposed that interoceptive sensitivity, particularly heartbeat, may be potentially involved in the transition from PA to PD [91, 92]. Interoception refers to the perception of the internal state of the body (e.g., heartbeat, respiration), while interoceptive sensitivity represents the degree of sensitivity to the internal sensations of the body and the accuracy of detection [93]. Given that PD patients often exhibit hypervigilance, this may be related to abnormalities in interoceptive sensitivity. An early cognitive model of panic proposed by Clark suggested that catastrophic misperceptions of somatic sensations lead to PAs [94]. Notably, based on the large body of clinical evidence finding that increased interoceptive sensitivity to the heartbeat is positively associated with PD, Domschke *et al.* focused specifically on and reviewed the prominent role of the heartbeat in this process [95]. He also pointed out that cardiac interoceptive sensitivity may serve as a potential intermediate phenotype of anxiety disorder to facilitate the understanding of complex diseases [95].

Neurotransmitter Systems in PD

Dysregulation of neurotransmitter systems, including serotonergic, glutamatergic, noradrenergic, and γ -aminobutyric systems, is associated with the fear network and physiopathology of PD [96]. Thus, determining the role of these neurotransmitters may contribute to the clinical management of PD.

The serotonergic system has been known to be involved in the pathogenesis and neurobiology of PD for decades, and two opposing hypotheses have been proposed: excessive or hyperactive 5-hydroxytryptamine (5-HT) [97, 98] and 5-HT deficiency or inactivity [99]. The main evidence for the first theory comes from drug trials. For instance, SSRIs have a biphasic effect when treating PD, ranging from an initial increase in anxiety and panic symptoms to a reduction after 2 weeks–3 weeks [100]. The first phase is due to the overactivation of receptors, while the second phase results from a progressive down-regulation of postsynaptic 5-HT receptor activity, compensated by chronic agonistic stimulation [101]. In addition, m-CPP, a direct HT agonist that increases 5-HT function by stimulating postsynaptic HT receptors, leads to increased anxiety and panic in PD patients, while having no significant effect on either normal individuals or patients with major depressive disorder [102].

Regarding the second hypothesis, Deakin *et al.* suggested that 5-HT in the dorsal raphe nucleus sends inhibitory projections to specific brain regions such as the PAG, and that panic ensues when this inhibitory input is reduced [99], suggesting that PD may be the result of 5-HT deficiency. Neuroimaging studies have provided additional evidence for this hypothesis. Maron *et al.* revealed lower 5-HT transporter binding in the temporal lobe, midbrain, and thalamus in PD patients than in healthy participants [103]. Another PET study focusing on central serotonin type-1A receptor binding found this to be significantly reduced in the brains of PD patients [104, 105].

In addition, γ -aminobutyric acid (GABA) is the most important inhibitory neurotransmitter in the CNS. Its receptors include GABA_A, GABA_B, and GABA_C, which are widely distributed in the brain [106]. GABA receptor dysfunction and/or downregulation of GABA concentration has been demonstrated in the brain of PD patients [106]. A previous PET-based study found an overall reduction in benzodiazepine site binding (on the GABA_A receptor) in the brains of PD patients [107]. Goddard *et al.* also demonstrated a markedly lower total occipital cortical GABA concentration in PD patients than in controls [108].

Accumulating evidence implies that the noradrenergic system may be involved in excessive arousal and abnormal ANS symptoms during PA [109, 110]. The α_2 -antagonist yohimbine activates noradrenergic neurons and causes increased plasma levels of 3-methoxy-4-hydroxyphenyl

glycol (a norepinephrine metabolite), triggering more severe somatic symptoms in PD patients [111]. In addition, genetic evidence suggests the presence of variants in genes associated with the noradrenergic system such as the norepinephrine transporter gene in PD patients [112].

The link between CCK-4 and PD has been widely demonstrated [77]. An fMRI study reported that a CCK-4 challenge may trigger PA by activating important components of the fear network, including the insular cortex, cingulate gyrus, and thalamic regions [113]. Furthermore, based on quantitative electroencephalography, Knott *et al.* reported the effects of continuous infusion of CCK-4 on hemispheric functional differences and found an increase in asymmetry and a decrease in coherence in slow-wave activity at the midtemporal recording site [114], leading to the speculation that dysfunction in the temporal cortex mediated by CCK-4 may be involved in PD.

Each of the above neurotransmitter systems has an important effect on PD, but they do not function independently, and the existence of interactions between them [66] suggests that PD is most likely the result of the dysregulation of a complex neurotransmitter network.

Pharmacotherapy and Psychotherapy for PD

Similar to other psychiatric disorders, PD treatment usually involves pharmacotherapy and psychotherapy [115]. Currently, four main classes of drugs are effective in treating PD: SSRIs/noradrenaline reuptake inhibitors, benzodiazepines, tricyclic antidepressants (TCAs), and monoamine oxidase inhibitors (MAOIs).

TCAs such as clomipramine and imipramine were previously used to treat PD [116, 117]. Although they can treat PD with promising efficacy, side-effects such as arrhythmia, constipation, sleep disturbance, and sexual dysfunction are also evident, so they are generally used as second-line drugs [118]. Given the key role of 5-HT in the pathogenesis of PD, SSRIs such as citalopram, escitalopram, and fluoxetine are currently used as first-line agents for the treatment of PD [119, 120]. Compared to TCAs, SSRIs are safer and more effective, have fewer side-effects, and are better tolerated [115]. Some antidepressants have a dual effect on noradrenergic and serotonergic neurons [121, 122].

The GABA system is deficient in PD patients [123, 124]. Thus, benzodiazepines such as alprazolam and diazepam, which are agonists of GABA_A benzodiazepine receptors (a complex containing chloride channels that are opened by agonists), may counteract this deficiency and ultimately have anxiolytic and sedative effects [125, 126]. However, the risks of developing tolerance and dependence, as well as sedation, lethargy, cognitive impairment, and motor coordination impairment, significantly limit the use of benzodiazepines [119, 127].

By blocking the catabolic effects of monoamine oxidase, MAOIs can increase brain levels of different types of neurotransmitters, including serotonin, norepinephrine, and dopamine, allowing them to continue to act on cells affected by depression [128]. However, MAOIs can have serious side-effects, including fatal hypertension after ingestion of foods with high levels of tyramine, and poor tolerability, both of which reduce overall patient compliance [129]. Therefore, they are not the first choice for PD treatment.

The clinical manifestations of PD are similar to an acute angina attack, acute ischemic attack, acute asthma attack, hyperthyroidism, and other diseases, so it can be easily misdiagnosed as cardiovascular or respiratory disease, which not only wastes medical resources and increases the economic burden on patients, but also causes great mental distress and loss of function in patients [130]. PD is also a comorbidity of multiple medical and psychiatric disorders, posing challenges for clinical intervention. Therefore, it is extremely important to strengthen the diagnosis, treatment, and prevention of PD. Factors such as effectiveness, side-effects, drug interactions, cost, and patient preferences need to be considered when developing specific treatment regimens. In the future, based on analysis of the neural circuits of PA, more specific targets should be explored to effectively treat PD and reduce side-effects.

CBT is recognized as the gold standard for the psychological treatment of anxiety disorders, and a large number of published meta-analyses have verified its excellent effectiveness for PD and other anxiety disorders [131–133]. CBT, along with SSRIs, has been the first-line treatment for PD with and without agoraphobia (PD/A) [134]. Actually, studies have reported that the effects of psychotherapy last longer than pharmacological therapy, and among the various psychotherapies, CBT is particularly recommended [135].

CBT consists of psychoeducation, cognitive restructuring, exposure therapy (imaginal, *in vivo*, and interoceptive), respiratory retraining, and other therapeutic components [136]. There are important differences in the efficacy and acceptability of these different components. For example, a meta-analysis has shown that the combination of exposure with relaxation training, and breathing retraining are most effective in treating PD/A [137]. In addition, Pompili *et al.* applied a component network meta-analysis to compare these different components in the treatment of PD and found greater advantages for interoceptive exposure (IE) and face-to-face therapy [136].

Notably, IE, a behavioral intervention that achieves treatment by gradually exposing patients to the physical sensations associated with panic, has long been recognized as an effective and primary therapy in the treatment of PD [138]. In this regard, Marcel van den Hout and his colleague, Eric Griez, played a seminal role, not only in the theoretical basis but also in clinical studies where they used repetitive

inhalation of CO₂ to reduce symptoms in PD patients [139–142].

Animal Studies

Animal-based studies, especially of mammals, have provided useful clues for the development of modern psychiatry [96]. Given evolutionary conservation, researchers have triggered fear-related behavior in animals to study the associated neural circuits, potentially providing important insights into PD in humans [85]. Therefore, in this section, we mainly summarize some major models that simulate panic-like responses in animals, based on which, the neural circuits that may be involved in PA are found.

Modeling PA in Rodents

Contemporary views suggest that the defensive reactions of animals in the face of an imminent threat resemble panic symptoms in humans [85]. Therefore, to better understand the neurobiology and pharmacology of PAs, many animal paradigms have been devised to assess panic-like responses induced by specific stimuli.

Considering that many aspects vary greatly among species, the judgment of animal model validity for human mental illness depends on three main criteria: face validity, postdictive and predictive validity, and construct validity [143]. Firstly, face validity refers to the symptoms in animal models that resemble the diagnostic criteria for clinically relevant diseases. For example, some PD patients typically feel panic and a desire to escape, which is reflected in explosive escape in mice and is an observable phenomenon with respect to PAs. In addition, abnormal ANS-related symptoms should also be detected in animal models. Postdictive validity indicates that panic-inducing procedures, such as CO₂ and sodium lactate, aggravate PA-associated symptoms in animal models, and therapeutic agents alleviate or eliminate these symptoms, while predictive validity means that the model can effectively predict the treatment effect of candidate drugs. Finally, construct validity is the degree of association between animal models and the pathophysiology underlying PAs [143].

CO₂ Exposure in Rodents

As discussed above, CO₂ inhalation is a valid model for triggering PAs in humans. However, the ability to study the neural circuitry of PD using human models is limited. CO₂ inhalation-induced panic in rodents is an alternative tool in this regard and significant progress has been made [30, 92].

Rodents cannot actively inhale CO₂ as humans do and considering that delivery of CO₂ using breathing masks

may be associated with stress [144], a preferable approach is to artificially expose mice to a compartment filled with a specific concentration of CO₂ for a period of time to induce PA while real-time behavioral assessments are performed. Experimental cages are usually made of clear Plexiglas, sealed with a removable cap, and connected *via* a flow valve to an air pump and a cylinder containing a mixture with a specific concentration of CO₂ [145].

When exposing mice to CO₂ for prolonged periods to assess panic-like behavior, the high concentrations of CO₂ used on humans are not feasible and have to be reduced appropriately. In addition, in humans, the subjective experience is a major component of the assessment and this cannot be assessed in rodents, which hinders the translation of data from animal studies to humans. In this regard, based on the pioneering work of Robert and Caroline Blanchard [146, 147], flight/avoidance behavior can be used as a reliable judgmental cue corresponding to panic and the desire to flee that some PD patients typically feel [145, 148]. While this is definitely true for some patients, there are also patients who experience a freezing response. Freezing in animals is considered a correlate of fear, thus the opposite response can also reflect a panic-like reaction [30]. In addition, similar physiological responses including increased blood pressure and respiratory rate are seen in both humans and animals, showing corresponding effects across species [30]. Thus, these metrics can be measured and incorporated in the future with the help of telemetry implants to further investigate the mechanisms.

Pharmacologically, both fluoxetine and imipramine affect panic-like responses during CO₂ exposure [145]. However, studies of this aspect are still scarce in rodents, and more evidence is needed to test the predictive validity of the model in the future. In terms of construct validity, the important role of acid-sensing ion channel 1a (ASIC1a) as a pH detector [149, 150] and orexin [151] as another candidate for panic states in animals and humans has been supported by many studies.

To conclude, compared to other animal models, it has great potential because it more closely resembles the way and mechanisms that trigger PAs in humans in clinical practice, and has been used to evaluate and screen novel anti-panic drugs [152].

Ultrasound-Induced Defensive Behaviors

Rats are known to use ultrasonic vocalizations to transmit information. The frequency and duration of the calls differ from environmental stimuli. Blanchard *et al.* found that rats retreat from the burrowing system and emit advanced ultrasonic calls at 18 kHz–24 kHz when exposed to a cat in an open area of the visible burrowing system [153]. Thereafter, Beckett *et al.* proposed a new model associated with

human panic by applying 22-kHz ultrasound (alarm calls) at an intensity of 65 dB to induce defensive behaviors in Lister-hooded (LH) rats [154].

Behavioral tests are performed in a test arena made of transparent Perspex walls and plastic floors that are placed in a soundproof box. During the test, rats are exposed to continuous ultrasound (22 kHz, 93 dB) for 1 min, followed by a 2-min rest. The distance traveled, percentage of time spent in hyperactive locomotion and freezing are recorded during this process.

Using this simple non-invasive model, Beckett *et al.* found increased neuronal activity in the dorsal PAG (dPAG), amygdala, paraventricular nucleus of the thalamus, and hypothalamus by assessing c-fos-like immunoreactivity [155]. Klein *et al.* found that ultrasound application induces hyperthermia and an increased heart rate in LH rats without altering blood corticosterone levels, thereby fulfilling the major physiological symptoms of human PAs [156]. Klein *et al.* also confirmed that the dorsolateral PAG (dlPAG) is involved in ultrasound-induced avoidance behavior, consistent with data obtained in humans [157]. Furthermore, pharmacological manipulation of ultrasound-triggered defensive behaviors in LH rats demonstrated the practical appeal of this PD model, as most compounds effective against PD (such as diazepam, alprazolam, and benzodiazepine) reduce escape responses [154, 158].

This alternative model for stimulating the dlPAG in animals to elicit panic-like responses is non-invasive and simple to manipulate [158]. However, different classes of anti-panic drugs selectively or non-selectively attenuate escape rather than freezing, whereas the nociceptin/orphanin FQ peptide receptor agonist Ro 64-6198 significantly affects freezing rather than escape, suggesting that these behaviors are regulated by different mechanisms [154].

Mouse Defense Test Battery (MDTB)

About three decades ago, Blanchard and colleagues developed the MDTB animal model following various pioneering psychiatric and pharmacological studies of defensive behavior in rodents [146, 159–161]. Similar to the elevated T-maze (ETM) test, the MDTB separates defense responses associated with GAD and PD.

The MDTB is conducted on an elliptical track with a total length of 6 m; the runway is connected by two straight sections and two curved sections separated by a wall. The entire apparatus is made of black Plexiglas, 0.80 m above the ground, allowing the experimenter to move the anesthetized rats at a uniform speed without the mice seeing the experimenter [162].

The MDTB assesses a range of behaviors exhibited by mice when faced with stimuli from a natural predator, the rat. During the experiment, a mouse is placed on the

elliptical track and an experimenter, wearing leather gloves, holds a deeply anesthetized rat and approaches at a fixed speed. Faced with the approaching rat, the mouse displays defense responses such as risk assessment, freezing, flight, and defensive attack. Based on pharmacological experiments, risk assessment, escape attempt, and defensive attack can be influenced by drugs used clinically to treat GAD, while flight represents a panic-like response [163]. For example, long-term injections of clinical anti-panic drugs, such as fluoxetine and alprazolam, significantly inhibit flight behavior in mice [160, 161, 164], while full agonists of the non-selective GABA_A-benzodiazepine receptor are less effective against flight, highlighting the limited utility of these drugs in treating clinical PD [159].

However, this model also has several shortcomings, including the need for anesthetized rats, trained observers, and sufficient space for the apparatus. The intensity of stimuli is also relatively low in anesthetized rats compared to other predators such as snakes. Notably, defense is stronger in the group exposed to the leather glove than in the group not exposed to any stimuli, indicating an effect on the responses of mice [165].

Elevated T-Maze

PAs have traditionally been considered a serious form of anxiety. With the separation of drug responses discovered by Klein [116] and the publication of DSM-III, PD was recognized as a separate entity from GAD. The ETM was developed in this general context, allowing the defense responses associated with GAD and PD to be tested separately in the same rat. Compared to the elevated plus-maze [166], the ETM has only three arms including one closed arm and two open arms with equal dimensions (50 cm × 12 cm) and 50 cm from the ground [148].

First, the rat is placed at the end of the enclosed arm and allowed to move freely. Out of a natural instinct to explore the new environment, the rat leaves the closed arm, during which the latency of the rat to poke its head out of the closed arm is recorded. Since the open and elevated arms are repulsive, the rat exhibits increased latency, i.e., acquisition of inhibitory avoidance, in three repetitions of the test. Subsequently, the rat is placed at the end of one open arm to perform a one-way escape task, while the latency for leaving the open arm generally does not change after three replicate experiments [148, 167].

Pharmacological studies have found that inhibitory avoidance relates to GAD and escape to PD. For example, the avoidance task is impaired by the acute administration of drugs clinically used to treat GAD such as diazepam, buspirone, and midazolam [148]. On the other hand, antidepressants affect one-way escape responses, consistent with the fact that these drugs have a therapeutic effect on PD [168,

169]. A recent study found that high doses of cocaine have a panicogenic-like effect that affects the escape behavior in rats during the ETM test and results in increased delta FosB immunoreactivity in the PAG and dorsal raphe nucleus [170]. Although the ETM appears to be a promising model for studying GAD and PD in rats, it is not stable in mice and needs further improvement [171].

Brain Mechanisms of PA

Amygdala

In 1989, Gorman *et al.* developed a hypothesis about the neuroanatomical basis of PD in an attempt to explain why both drugs and CBT are effective in treating PD [172]. A decade later, they revised the hypothesis based on advances in the neuroanatomy of conditioned fear in animals, emphasizing the importance of the amygdala in panic [96]. Currently, it has been established that the amygdala-centered fear network integrates sensory inputs from other brain regions to coordinate animal defense behavior and is associated with psychiatric disorders such as PD in humans [173].

In addition, ASIC1a detects a decrease in extracellular pH; it is expressed at particularly high levels in the amygdala [174, 175], and its relationship with PD has been studied in animal models of PAs induced by CO₂ inhalation. Inhaled CO₂ interferes with pH homeostasis and a lowered pH activates ASIC1a channels; the amygdala thereby plays a chemosensory role to elicit fear behavior [175]. The study showed that in *ASIC-/-* mice, the CO₂-induced fear response is eliminated, and local injection of AAV-*ASIC1a* in the amygdala rescues this [149]. Furthermore, microinjection of acidified artificial cerebral spinal fluid (ACSF) into the amygdala of wild-type mice results in a decrease in local pH and elicits a fear response similar to that induced by CO₂ [174]. A recent study found that mice with a damaged amygdala exhibit more frequent jumping behavior when exposed to 10% CO₂, and further experiments demonstrated that the amygdala plays a dual role in CO₂-induced defense responses [176].

Midbrain

The midbrain is a complex and heterogeneous nucleus that mediates many aspects of predation, defense responses to threat, reward, locomotion, and stress adaptation [177]. Here, we focus on the role of the superior colliculus (SC) and PAG in defense responses in mammals given the large body of literature on these two evolutionarily well-conserved brain regions and that defense responses triggered by an imminent threat in mammals can be linked to PAs in humans.

Superior Colliculus

When facing aerial predators, only vision provides useful information, and the SC plays a key role as a retinal receptor structure in vision-evoked defensive behavior [178]. Visual signals of overhead threats can be classified into two categories: sweeping and looming [5]. The former, representing a potential and distant threat, activates a class of SC neurons (“wide-field cells”) [179], while the latter, representing an approaching predator and an imminent threat, induces escape behavior [180]. The involvement of the SC in defense responses to imminent stimuli has been studied in different species, including *Drosophila* [181], zebrafish [182, 183], mice [184], and cats [185].

Substantial evidence has shown that the SC receives retinal ganglion cell input and then projects it to multiple brain regions to mediate defensive behavior in a bottom-up manner. Evans *et al.* revealed that excitatory neuronal activity in the deep medial SC represents the salience of threatening stimuli, initiating escape through synaptic thresholds at the dPAG level [186]. Shang *et al.* revealed a “retina-SC-parabigeminal nucleus (PBGN)-amygdala-hypothalamus” pathway mediating the vision-evoked fear response, with the SC PV⁺ neuron population as a key mediator [187]. However, in parallel, Wei *et al.* proposed a new “medial region of the SC intermediate layer-lateral posterior nucleus (LPTN)-lateral amygdala” pathway involved in innate defense responses to visual threats [188]. To clarify how different defensive behaviors (freezing and fleeing) are selected, Shang *et al.* systematically investigated the role of SC PV⁺ neurons and their downstream pathways and identified two groups of SC PV⁺ neurons projecting to the LPTN and PBGN, respectively, forming different visual pathways to coordinate dimorphic fear behaviors, which compete with each other and produce a winner-takes-all behavioral outcome [189].

Xie *et al.* recently demonstrated diversity in SC neuronal subtypes and found that LPTN-projecting optic nerve layer neurons highly express the *Cbln2* marker. Optogenetic activation of the Cbln2⁺ SC-LPTN pathway selectively triggers escape behavior [190]. Top-down cortical inputs to the SC are also involved in behavioral responses to visual cues in mice, providing a potential neural substrate for the regulation of defense responses [191, 192]. Taken together, the above studies demonstrate that the SC is a key structure in the production of defense responses in mammals and thus may be involved in human PAs.

Periaqueductal Gray Matter

At present, there is sufficient preclinical and clinical evidence linking the PAG, especially the dPAG, to the etiology of PAs. In mammals, evidence suggests that the PAG is the most important component of neural circuits that regulate

defense responses in the face of an imminent threat [193, 194]. Deng *et al.* found that activation of dPAG neurons triggers behaviors such as escape and conditioned avoidance and subsequently identified two classes of neurons: assessment and flight cells [195]. A recent study also reported these two groups of neurons in the dPAG and found that flight-positive cells function as initiators encoding motor patterns [196].

In addition to receiving input from the SC, which enables visual signals to trigger defense responses [189], the PAG also receives signals from many cortical and subcortical areas, thereby integrating other types of threat-related information [197, 198]. The PAG receives information about predator cues, mainly pheromones and olfactory information, from the medial amygdala (MeA) via the ventromedial hypothalamus (VMH) to initiate the corresponding defense behavior [194, 199, 200]. The PAG also receives input from the lateral hypothalamus, and activation of PAG-projecting lateral hypothalamus glutamatergic neurons leads to escape and jumping, while inhibition increases the escape threshold [201]. The auditory cortex-inferior colliculus-PAG pathway drives sound-induced innate defensive behavior [202]. Similar phenomena occur in other PAG-projecting structures, such as the zona incerta, a major subthalamic structure that bidirectionally regulates the dynamics of active defensive behavior [203].

The dPAG also sends projections to multiple brain regions including the mesencephalic motor area (MLR), which controls high-speed movement during escape [204, 205]. Abnormalities in dPAG projections to the amygdala may be involved in fear-related psychiatric disorders, such as PD and post-traumatic stress disorder [206].

In several studies, dPAG aversive stimuli have been combined with self-interruption responses to test the effects of anti-panic drugs [207]. Jenck *et al.* showed that anti-panic drugs (alprazolam and clonazepam) reduce the aversive effects of dPAG stimulation while panic-inducing complexes (yohimbine and caffeine) have the opposite effect [208]. In addition, drugs that treat PD by increasing the action of serotonin are also able to reduce the panic-like behavior induced by stimulating the PAG in mice [209].

In fact, early studies have identified an important role for 5-HT in the PAG in animal defense behavior [210, 211]. At that time, most psychiatrists considered anxiety to be a single emotion that could be alleviated with anxiolytic drugs. Some studies supported the notion that 5-HT acts on the forebrain to exert anxiogenic effects, but the effects of 5-HT in the PAG are inconsistent [212]. Other studies that treated rats with 5-hydroxytryptophan and the serotonin reuptake inhibitor clomipramine [213] or with electrical stimulation of the ACC [214] reported a reduction in switch-off responding (dPAG-evoked responses in rats considered a model of anxiety at the time), demonstrating that 5-HT reduces anxiety.

In the following decades, the concept of the brain defense system emerged [215], in which defense strategies were divided into three levels, corresponding to different brain regions [7]. As a result, PD began to be recognized as a diagnostic entity distinct from other psychiatric disorders such as GAD. Finally, based on many studies, 5-HT was considered to regulate GAD and PD in opposite directions, with the promoting effect of 5-HT on forebrain limbic structures related to GAD and the inhibiting effect of 5-HT on the PAG related to PD [216]. This hypothesis was later verified through cumulative experiments using the ETM panic model, which also revealed that 5-HT_{1A} and 5-HT_{2A} receptors may mediate the anti-stunning effects of 5-HT in the dPAG and must be functional to activate their expression [217].

Hypothalamus

The hypothalamus is an evolutionarily conserved structure that plays an important role in defensive behaviors. Early work by Hess and colleagues laid the foundation for the view that the hypothalamus is a key part of coordinated panic defense responses [218, 219]. Along with other studies, the concept of the medial hypothalamic area (MHZ) was developed, including the dorsomedial part of the VMH (VMHdm), AHN, and dorsal premammillary nucleus (PMD) [220–222].

The VMHdm is thought to occupy a central position in the MHZ defense system, receiving direct inputs of threat-related information from the cortex and MeA and projecting to the PAG to elicit defense responses [223, 224]. Optogenetic activation of the VMHdm in mice interrupts ongoing activity and induces freezing and escape behavior [225, 226]. Silva *et al.* reported that exposure to predators significantly increases c-Fos expression in the VMHdm [199, 227].

Similarly, Kunwar *et al.* reported that direct activation of VMHdm SF1 neurons is sufficient to evoke a variety of defensive behaviors, with weaker stimuli inducing freezing and stronger stimuli evoking bursts of activity, and specific ablation weakening defensive behaviors in a variety of contexts. Kunwar *et al.* also proposed that the hypothalamus does not simply act as a relay station for amygdala output, but is itself involved in threat arousal or a defensive motivational state, which may be associated with the human emotion of “fear” or “panic” [228]. In addition, the projection of the VMHdm to the ventrolateral anastomosis of the rostral ventrolateral medulla (RVLM) may be involved in ANS-related responses such as tachycardia and hyperventilation [229].

The PMD also occupies an important position in the MHZ system, providing the most intensive input to the panicogenic PAG [198]. Staples *et al.* found that the PMD is part of the hypothalamus that responds most strongly to

environments previously associated with cat odors [230]. Similarly, Canteras *et al.* showed that the PMD mediates the expression of unconditioned defense responses [231, 232]. Moreover, in complex experimental scenarios, Wang *et al.* reported that the PMD recruits spatial navigation networks and spatial escape induction circuits and integrates both functions to produce context-specific escape [233]. Notably, they also reported that chemogenetic inhibition and excitation of PMD-cck neurons decreases and increases panic-related escape attempts in mice, respectively, where jumping escape is triggered by high-intensity threats such as CO₂.

Another important component of the MHZ is the AHN. Paschoalin-Maurin *et al.* found that when confronted with a venomous coral snake, hamsters exhibit panic-like behavioral responses with dramatically increased c-Fos expression in the AHN [234]. Xie *et al.* recently revealed that inhibitory neurons in the AHN encode threat-related sensory information and specifically modulate defensive aggression through neural circuits projecting to the ventrolateral PAG [6].

Given the role of the rodent hypothalamus in the “fight or flight” reaction [235, 236], Shekhar and colleagues established a rat model of PD by unilaterally infusing 1-allylglycine (a GABA synthesis inhibitor) into the dorsomedial/perifornical hypothalamic (DMH/PeF) region [237]. Notably, panic-prone rats exhibit panic-associated behaviors and cardiorespiratory responses after administration of 0.5 M NaLac, yohimbine, and CO₂ [238, 239]. Research has also shown that panic-prone rats respond positively to the benzodiazepine alprazolam [240].

Accumulating evidence suggests that the balance between GABA inhibition and glutamate excitation in the DMH/PeF is related to a panic-like response [237, 241, 242]. Glutamatergic receptors may represent a novel group of targets for the treatment of PD. For example, pretreatment of panic-prone rats with a selective mGluR2 agonist (CBiPES or THIC) attenuates panic-like responses [237]. Likewise, LY354740, a selective group II mGlu receptor agonist, blocks lactate-induced panic-like responses in DMH GABA-dysfunctional rats [243, 244].

Orexin (hypocretin) (ORX/HCRT), first described in 1998 [245], is a neuropeptide that is highly enriched in the hypothalamus of rodents [245, 246] and humans [247]. ORX neurons broadly project to almost every part of the CNS and play a critical role in controlling behavioral states, appetite, and autonomic functions [245, 246, 248, 249]. Johnson *et al.* demonstrated that deletion of *preproORX* mRNA, a product of the hypothalamic ORX gene (*Hcrt*), or administration of ORX1 receptor antagonists block panic-like responses in rats [250]. Similarly, ORX1 receptor antagonists attenuate the panic state in rats [151]. These findings indicate that abnormal activation of ORX neurons may be associated with the formation of a panic-prone state in animal models. In

conclusion, the hypothalamus as a “defense area” plays a considerable role in the pathogenesis of PA.

Other Critical Structures

In addition to the well-studied major brain regions, other structures have also received attention.

The VTA is a heterogeneous nucleus containing dopamine, aminobutyric acid, and glutamate neurons and is associated with reward and aversion [251, 252]. However, growing evidence also suggests that the VTA plays a role in innate fear responses, such as the acquisition of conditioned fear [253, 254] and the processing of predator-related information [255]. Furthermore, VTA-GABA neurons receiving input from lateral hypothalamus-glutamate neurons mediate place avoidance [256]. Zhou *et al.* also found that VTA-GABA neurons receiving SC excitatory inputs and projecting to the CeA are involved in looming-evoked fly-to-nest behavior [257]. A more recent study has shown that VTA-glutamate neurons mediate security by encoding innate escape responses, unlike VTA-GABA neurons, by facilitating concealment behavior [258].

Locomotion is another important component of defensive behavior and changing speed in response to behavioral requirements is necessary [259]. In many species, the MLR, a structure that anatomically overlaps with the pedunculopontine nucleus, cuneiform nucleus, and mesencephalic reticular nucleus in mammals, is critical for locomotion [260]. Activation of glutamatergic MLR neurons induces locomotion at short latencies, whereas photo-inhibition impairs movement [261, 262]. In contrast to glutamatergic neurons, optogenetic activation of MLR GABA neurons leads to a motor arrest, and these neurons receive dense inputs mainly from limbic regions, suggesting that MLR GABA neurons are associated with fear-related behaviors [261, 263].

In addition, regarding the mPFC, Halladay and Blair recorded the neural activity of the mPFC and dPAG during two defense behaviors provoked by a fear-conditioned auditory CS in rats and suggested that the mPFC may contribute to the regulation of defense strategies through the projection of the PAG [264]. Another study showed that activation of the mPFC-dPAG pathway is sufficient to drive both place avoidance and defensive behavior [265]. Pre-treatment with N-methyl-D-aspartate receptor antagonism in the prelimbic division of the mPFC in rats has been found to reduce panic-like behavior, implying a critical role for the mPFC [266].

Research on fear memory has contributed to the understanding of various anxiety disorders, and much of the research has focused on the hippocampus, which has long been thought to be the center of learning and memory [267]. Using predator odor as an unconditioned stimulus, it has

been found that both the dorsal (DH) and ventral hippocampus (VH) are involved in contextual fear learning, but with different functions. The VH is more associated with the fear response to predator odor, whereas the DH is mainly involved in spatial representation [268].

The LC has traditionally been associated with sleep/wake and attention [269–271]. Given the high attention span of an animal when confronted with a predator, the LC may be involved in this process. Accordingly, activation of TH neurons in the LC accelerates the defense response to looming stimuli in mice [272]. An air-puff elicits a defensive freezing response in mammals accompanied by LC activation [273].

Taken together, the PA is a complicated abnormal physiological state, which involves interactions between multiple brain regions (Fig. 3). The above-mentioned brain regions may be associated not only with PD but also with many other psychiatric disorders. For example, it has been reported that the amygdala, ACC, and hippocampus may be involved in all anxiety disorders [274]. Of note, electrical stimulation of the DPAG [275] and certain hypothalamic areas [276] in humans induces clinical panic attack-like symptoms, suggesting that these two brain regions may more directly and specifically regulate PD. Therefore, more studies are needed in the future to further confirm the specific roles played by these brain regions.

Conclusions

In this review, we summarize the current information on PD/PAs derived from animal and human studies. Although significant progress has been made in the elucidation of the mechanisms and treatment of PD, many questions remain to

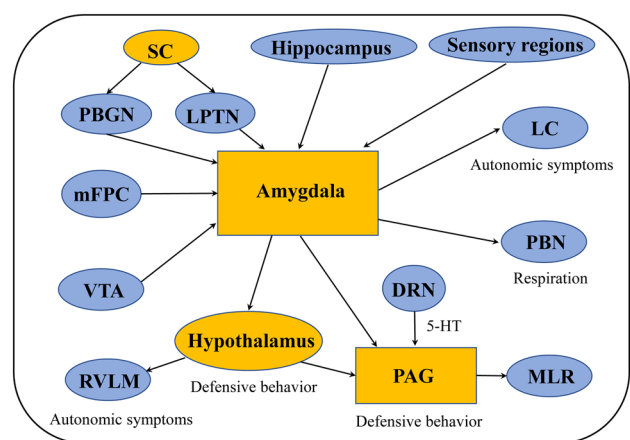


Fig. 3 Overview of the possible neural circuits underlying panic attacks. The hypothalamus and PAG are primarily associated with the expression of panic-related defensive behaviors. The LC and RVLM are involved in autonomic nervous system symptoms. The PBN regulates respiration.

be addressed. In humans, given the heterogeneity and complexity of PD, more advanced techniques and more detailed diagnostic criteria for further typing of PD are needed to treat different PD patients more precisely. And the establishment of animal PA models has greatly contributed to the in-depth elucidation of the pathogenesis of PD. However, considering that behavioral performance in animals and verbal self-reports in humans are not interchangeable, the exact differences between these two states are poorly understood and further characterization is needed. Ultimately, advances in the basic neuroscience of PAs are of great importance and contribute to breakthroughs in research on clinical psychiatric disorders.

Conflict of interest The authors declare no conflict of interests.

References

1. American Psychiatric Association (1980) Diagnostic and statistical manual of mental disorders, 3rd edn. American Psychiatric Association, Washington, DC, pp 230–232.
2. Vahia VN. Diagnostic and statistical manual of mental disorders 5: A quick glance. *Indian J Psychiatry* 2013, 55: 220–223.
3. Livermore JJA, Klaassen FH, Bramson B, Hulsman AM, Meijer SW, Held L. Approach-avoidance decisions under threat: The role of autonomic psychophysiological states. *Front Neurosci* 2021, 15: 621517.
4. Gowda SBM, Banu A, Salim S, Peker KA, Mohammad F. Serotonin distinctly controls behavioral states in restrained and freely moving *Drosophila*. *iScience* 2023, 26: 105886.
5. De Franceschi G, Vivattanasarn T, Saleem AB, Solomon SG. Vision guides selection of freeze or flight defense strategies in mice. *Curr Biol* 2016, 26: 2150–2154.
6. Xie Z, Gu H, Huang M, Cheng X, Shang C, Tao T, *et al.* Mechanically evoked defensive attack is controlled by GABAergic neurons in the anterior hypothalamic nucleus. *Nat Neurosci* 2022, 25: 72–85.
7. Blanchard RJ, Flannelly KJ, Blanchard DC. Defensive behavior of laboratory and wild *Rattus norvegicus*. *J Comp Psychol* 1986, 100: 101–107.
8. Strawn JR, Geraciotti L, Rajdev N, Clemenza K, Levine A. Pharmacotherapy for generalized anxiety disorder in adult and pediatric patients: An evidence-based treatment review. *Expert Opin Pharmacother* 2018, 19: 1057–1070.
9. Eaton WW, Bienvenu OJ, Miloyan B. Specific phobias. *Lancet Psychiatry* 2018, 5: 678–686.
10. Caldirola D, Alciati A, Cuniberti F, Perna G. Experimental drugs for panic disorder: An updated systematic review. *J Exp Pharmacol* 2021, 13: 441–459.
11. Graeff FG. Neuroanatomy and neurotransmitter regulation of defensive behaviors and related emotions in mammals. *Braz J Med Biol Res* 1994, 27: 811–829.
12. Xu P, Chen A, Li Y, Xing X, Lu H. Medial prefrontal cortex in neurological diseases. *Physiol Genomics* 2019, 51: 432–442.
13. Mobbs D, Petrovic P, Marchant JL, Hassabis D, Weiskopf N, Seymour B, *et al.* When fear is near: Threat imminence elicits prefrontal-periaqueductal gray shifts in humans. *Science* 2007, 317: 1079–1083.
14. De Oca BM, DeCola JP, Maren S, Fanselow MS. Distinct regions of the periaqueductal gray are involved in the acquisition and expression of defensive responses. *J Neurosci* 1998, 18: 3426–3432.
15. McNaughton N, Corr PJ. A two-dimensional neuropsychology of defense: Fear/anxiety and defensive distance. *Neurosci Biobehav Rev* 2004, 28: 285–305.
16. Maren S. Neuroscience. The threatened brain. *Science* 2007, 317: 1043–1044.
17. Caldirola D, Perna G. Toward a personalized therapy for panic disorder: Preliminary considerations from a work in progress. *Neuropsychiatr Dis Treat* 2019, 15: 1957–1970.
18. Zaubler TS, Katon W. Panic disorder in the general medical setting. *J Psychosom Res* 1998, 44: 25–42.
19. Aust G. Anxiety syndrome in vertigo patients. A comparison of neuro-otologic findings in patients with and without anxiety. *Laryngorhinootologie* 1995, 74: 601–605.
20. Klein DF. False suffocation alarms, spontaneous panics, and related conditions. An integrative hypothesis. *Arch Gen Psychiatry* 1993, 50: 306–317.
21. Briggs AC, Stretch DD, Brandon S. Subtyping of panic disorder by symptom profile. *Br J Psychiatry* 1993, 163: 201–209.
22. Okuro RT, Freire RC, Zin WA, Quagliato LA, Nardi AE. Panic disorder respiratory subtype: Psychopathology and challenge tests—an update. *Braz J Psychiatry* 2020, 42: 420–430.
23. Stein DJ. The respiratory subtype of panic disorder: Reflections on the past and future of biological psychiatry. *Braz J Psychiatry* 2020, 42: 340–341.
24. Nardi AE, Valença AM, Nascimento I, Lopes FL, Mezzasalma MA, Freire RC, *et al.* A three-year follow-up study of patients with the respiratory subtype of panic disorder after treatment with clonazepam. *Psychiatry Res* 2005, 137: 61–70.
25. Yoon HK, Kang J, Kwon DY, Ham BJ. Frontoparietal cortical thinning in respiratory-type panic disorder: A preliminary report. *Psychiatry Investig* 2016, 13: 146–151.
26. Freire RC, Perna G, Nardi AE. Panic disorder respiratory subtype: Psychopathology, laboratory challenge tests, and response to treatment. *Harv Rev Psychiatry* 2010, 18: 220–229.
27. Onur E, Alkin T, Tural U. Panic disorder subtypes: Further clinical differences. *Depress Anxiety* 2007, 24: 479–486.
28. Nardi AE, Nascimento I, Valença AM, Lopes FL, Mezzasalma MA, Zin WA, *et al.* Respiratory panic disorder subtype: Acute and long-term response to nortriptyline, a noradrenergic tricyclic antidepressant. *Psychiatry Res* 2003, 120: 283–293.
29. Guttmacher LB, Murphy DL, Insel TR. Pharmacologic models of anxiety. *Compr Psychiatry* 1983, 24: 312–326.
30. Leibold NK, van den Hove DL, Viechtbauer W, Buchanan GF, Goossens L, Lange I, *et al.* CO₂ exposure as translational cross-species experimental model for panic. *Transl Psychiatry* 2016, 6: e885.
31. Rassovsky Y, Kushner MG. Carbon dioxide in the study of panic disorder: Issues of definition, methodology, and outcome. *J Anxiety Disord* 2003, 17: 1–32.
32. Drury AN. The percentage of carbon dioxide in the alveolar air, and the tolerance to accumulating carbon dioxide in case of so-called “irritable heart.” *Heart* 1918, 4(7): 165–173.
33. Cohen ME, White PD. Life situations, emotions, and neurocirculatory asthenia (anxiety neurosis, neurasthenia, effort syndrome). *Psychosom Med* 1951, 13: 335–357.
34. Amaral JM, Spadaro PT, Pereira VM, Silva AC, Nardi AE. The carbon dioxide challenge test in panic disorder: A systematic review of preclinical and clinical research. *Braz J Psychiatry* 2013, 35: 318–331.
35. Bailey JE, Kendrick A, Diaper A, Potokar JP, Nutt DJ. A validation of the 7.5% CO₂ model of GAD using paroxetine and lorazepam in healthy volunteers. *J Psychopharmacol* 2007, 21: 42–49.

36. Perna G, Bertani A, Arancio C, Ronchi P, Bellodi L. Laboratory response of patients with panic and obsessive-compulsive disorders to 35% CO₂ challenges. *Am J Psychiatry* 1995, 152: 85–89.
37. Perna G, Cocchi S, Politi E, Bellodi L. CO₂ challenge in nonclinical subjects. *Am J Psychiatry* 1997, 154: 886–887.
38. Perna G, Battaglia M, Garberi A, Arancio C, Bertani A, Bellodi L. Carbon dioxide/oxygen challenge test in panic disorder. *Psychiatry Res* 1994, 52: 159–171.
39. Perna G, Cocchi S, Allevi L, Bussi R, Bellodi L. A long-term prospective evaluation of first-degree relatives of panic patients who underwent the 35% CO₂ challenge. *Biol Psychiatry* 1999, 45: 365–367.
40. Perna G, Bussi R, Allevi L, Bellodi L. Sensitivity to 35% carbon dioxide in patients with generalized anxiety disorder. *J Clin Psychiatry* 1999, 60: 379–384.
41. Schruers K, Klaassen T, Pols H, Overbeek T, Deutz NE, Griez E. Effects of tryptophan depletion on carbon dioxide provoked panic in panic disorder patients. *Psychiatry Res* 2000, 93: 179–187.
42. Schruers K, van Diest R, Nicolson N, Griez E. L-5-Hydroxytryptophan induced increase in salivary cortisol in panic disorder patients and healthy volunteers. *Psychopharmacology* 2002, 161: 365–369.
43. Schruers K, Griez E. The effects of tianeptine or paroxetine on 35% CO₂ provoked panic in panic disorder. *J Psychopharmacol* 2004, 18: 553–558.
44. van Duinen MA, Schruers KR, Jaegers E, Maes M, Griez EJ. Hypothalamic-pituitary-adrenal axis function following a 35% CO₂ inhalation in healthy volunteers. *Prog Neuropsychopharmacol Biol Psychiatry* 2004, 28: 279–283.
45. Cai M, Wang H, Song H, Yang R, Wang L, Xue X, *et al.* Lactate is answerable for brain function and treating brain diseases: Energy substrates and signal molecule. *Front Nutr* 2022, 9: 800901.
46. Rabinowitz JD, Enerbäck S. Lactate: The ugly duckling of energy metabolism. *Nat Metab* 2020, 2: 566–571.
47. Pitts FN Jr, McClure JN Jr. Lactate metabolism in anxiety neurosis. *N Engl J Med* 1967, 277: 1329–1336. <https://doi.org/10.1056/NEJM196712212772502>.
48. Kellner M. Experimental panic provocation in healthy man—A translational role in anti-panic drug development? *Dialogues Clin Neurosci* 2011, 13: 485–493.
49. Wiese AD, Boutros NN. Diagnostic utility of sodium lactate infusion and CO₂-35% inhalation for panic disorder. *Neuropsychobiology* 2019, 78: 59–69.
50. Liebowitz MR, Gorman JM, Fyer AJ, Levitt M, Dillon D, Levy G, *et al.* Lactate provocation of panic attacks. II. Biochemical and physiological findings. *Arch Gen Psychiatry* 1985, 42: 709–719.
51. Peskind ER, Jensen CF, Pascualy M, Tsuang D, Cowley D, Martin DC, *et al.* Sodium lactate and hypertonic sodium chloride induce equivalent panic incidence, panic symptoms, and hypernatremia in panic disorder. *Biol Psychiatry* 1998, 44: 1007–1016.
52. Magarian GJ. Hyperventilation syndromes: Infrequently recognized common expressions of anxiety and stress. *Medicine (Baltimore)* 1982, 61: 219–236.
53. Hibbert G, Pilsbury D. Hyperventilation: Is it a cause of panic attacks? *Br J Psychiatry* 1989, 155: 805–809.
54. Meuret AE, Ritz T, Wilhelm FH, Roth WT. Voluntary hyperventilation in the treatment of panic disorder—Functions of hyperventilation, their implications for breathing training, and recommendations for standardization. *Clin Psychol Rev* 2005, 25: 285–306.
55. Gardner WN. The pathophysiology of hyperventilation disorders. *Chest* 1996, 109: 516–534.
56. Nardi AE, Valença AM, Nascimento I, Mezzasalma MA, Zin WA. Hyperventilation in panic disorder and social phobia. *Psychopathology* 2001, 34: 123–127.
57. Nardi AE, Valença AM, Nascimento I, Zin WA. Panic disorder and obsessive compulsive disorder in a hyperventilation challenge test. *J Affect Disord* 2002, 68: 335–340.
58. Nardi AE, Nascimento I, Valença AM, Lopes FL, Zin WA, Mezzasalma MA, *et al.* A breath-holding challenge in panic disorder patients, their healthy first-degree relatives, and normal controls. *Respir Physiol Neurobiol* 2002, 133: 43–47.
59. Nardi AE, Valença AM, Nascimento I, Zin WA. Hyperventilation challenge test in panic disorder and depression with panic attacks. *Psychiatry Res* 2001, 105: 57–65.
60. Nardi AE, Valença AM, Mezzasalma MA, Levy SP, Lopes FL, Nascimento I, *et al.* Comparison between hyperventilation and breath-holding in panic disorder: Patients responsive and non-responsive to both tests. *Psychiatry Res* 2006, 142: 201–208.
61. Nardi AE, Lopes FL, Valença AM, Nascimento I, Mezzasalma MA, Zin WA. Psychopathological description of hyperventilation-induced panic attacks: A comparison with spontaneous panic attacks. *Psychopathology* 2004, 37: 29–35.
62. Freire RC, Lopes FL, Valença AM, Nascimento I, Veras AB, Mezzasalma MA, *et al.* Panic disorder respiratory subtype: A comparison between responses to hyperventilation and CO₂ challenge tests. *Psychiatry Res* 2008, 157: 307–310.
63. Gorman JM, Fyer MR, Goetz R, Askanazi J, Liebowitz MR, Fyer AJ, *et al.* Ventilatory physiology of patients with panic disorder. *Arch Gen Psychiatry* 1988, 45: 31–39.
64. Ivy AC, Oldberg E. A hormone mechanism for gall-bladder contraction and evacuation. *Am J Physiol Leg Content* 1928, 86: 599–613.
65. Rehfeld JF. Cholecystokinin and the hormone concept. *Endocr Connect* 2021, 10: R139–R150.
66. Rehfeld JF. Cholecystokinin and panic disorder: Reflections on the history and some unsolved questions. *Molecules* 2021, 26: 5657.
67. Bradwejn J, de Montigny C. Benzodiazepines antagonize cholecystokinin-induced activation of rat hippocampal neurones. *Nature* 1984, 312: 363–364.
68. Bradwejn J, Koszycki D. Cholecystokinin and panic disorder: Past and future clinical research strategies. *Scand J Clin Lab Invest Suppl* 2001, 234: 19–27.
69. Bradwejn J, Koszycki D, Couëtoux du Tertre A, van Megen H, den Boer J, Westenberg H. The panicogenic effects of cholecystokinin-tetrapeptide are antagonized by L-365, 260, a central cholecystokinin receptor antagonist, in patients with panic disorder. *Arch Gen Psychiatry* 1994, 51: 486–493.
70. Bradwejn J, Koszycki D, Bourin M. Dose ranging study of the effects of cholecystokinin in healthy volunteers. *J Psychiatry Neurosci* 1991, 16: 91–95.
71. Bradwejn J, Koszycki D, Shriqui C. Enhanced sensitivity to cholecystokinin tetrapeptide in panic disorder. Clinical and behavioral findings. *Arch Gen Psychiatry* 1991, 48: 603–610.
72. Bradwejn J, Koszycki D. Imipramine antagonism of the panicogenic effects of cholecystokinin tetrapeptide in panic disorder patients. *Am J Psychiatry* 1994, 151: 261–263.
73. van Megen HJGM, Westenberg HGM, den Boer JA, Slaap B, Scheepmakers A. Effect of the selective serotonin reuptake inhibitor fluvoxamine on CCK-4 induced panic attacks. *Psychopharmacology* 1997, 129: 357–364.
74. Shlik J, Aluoja A, Vasar V, Vasar E, Podar T, Bradwejn J. Effects of citalopram treatment on behavioural, cardiovascular and neuroendocrine response to cholecystokinin tetrapeptide challenge in patients with panic disorder. *J Psychiatry Neurosci* 1997, 22: 332–340.
75. Javanmard M, Shlik J, Kennedy SH, Vaccarino FJ, Houle S, Bradwejn J. Neuroanatomic correlates of CCK-4-induced panic attacks in healthy humans: A comparison of two time points. *Biol Psychiatry* 1999, 45: 872–882.

76. Eser D, Leicht G, Lutz J, Wenninger S, Kirsch V, Schüle C, *et al.* Functional neuroanatomy of CCK-4-induced panic attacks in healthy volunteers. *Hum Brain Mapp* 2009, 30: 511–522.
77. Holzsneider K, Mulert C. Neuroimaging in anxiety disorders. *Dialogues Clin Neurosci* 2011, 13: 453–461.
78. Eser D, Schüle C, Baghai T, Floesser A, Krebs-Brown A, Enunwa M, *et al.* Evaluation of the CCK-4 model as a challenge paradigm in a population of healthy volunteers within a proof-of-concept study. *Psychopharmacology* 2007, 192: 479–487.
79. Olson VG, Rockett HR, Reh RK, Redila VA, Tran PM, Venkov HA, *et al.* The role of norepinephrine in differential response to stress in an animal model of posttraumatic stress disorder. *Biol Psychiatry* 2011, 70: 441–448.
80. Sperl MFJ, Panitz C, Skoluda N, Nater UM, Pizzagalli DA, Hermann C, *et al.* Alpha-2 adrenoreceptor antagonist yohimbine potentiates consolidation of conditioned fear. *Int J Neuropsychopharmacol* 2022, 25: 759–773.
81. Kaplan JS, Arnkoff DB, Glass CR, Tinsley R, Geraci M, Hernandez E, *et al.* Avoidant coping in panic disorder: A yohimbine biological challenge study. *Anxiety Stress Coping* 2012, 25: 425–442.
82. Klein E, Zohar J, Geraci MF, Murphy DL, Uhde TW. Anxiogenic effects of m-CPP in patients with panic disorder: Comparison to caffeine's anxiogenic effects. *Biol Psychiatry* 1991, 30: 973–984.
83. Gutman DA, Coplan J, Papp L, Martinez J, Gorman J. Doxapram-induced panic attacks and cortisol elevation. *Psychiatry Res* 2005, 133: 253–261.
84. Garakani A, Buchsbaum MS, Newmark RE, Goodman C, Aaronson CJ, Martinez JM, *et al.* The effect of doxapram on brain imaging in patients with panic disorder. *Eur Neuropsychopharmacol* 2007, 17: 672–686.
85. Dresler T, Guhn A, Tupak SV, Ehli AC, Herrmann MJ, Fallgatter AJ, *et al.* Revise the revised? New dimensions of the neuroanatomical hypothesis of panic disorder. *J Neural Transm* 2013, 120: 3–29.
86. Preter M, Klein DF. Panic, suffocation false alarms, separation anxiety and endogenous opioids. *Prog Neuropsychopharmacol Biol Psychiatry* 2008, 32: 603–612.
87. Vickers K, McNally RJ. Respiratory symptoms and panic in the National Comorbidity Survey: A test of Klein's suffocation false alarm theory. *Behav Res Ther* 2005, 43: 1011–1018.
88. Eysenck HJ. The conditioning model of neurosis. *Behav Brain Sci* 1979, 2: 155–166.
89. Bouton ME, Mineka S, Barlow DH. A modern learning theory perspective on the etiology of panic disorder. *Psychol Rev* 2001, 108: 4–32.
90. Kyriakoulis P, Kyrios M. Biological and cognitive theories explaining panic disorder: A narrative review. *Front Psychiatry* 2023, 14: 957515.
91. Yoris A, Esteves S, Couto B, Melloni M, Kichic R, Cetkovich M, *et al.* The roles of interoceptive sensitivity and metacognitive interoception in panic. *Behav Brain Funct* 2015, 11: 14.
92. Leibold NK, van den Hove DLA, Esquivel G, De Cort K, Goossens L, Strackx E, *et al.* The brain acid-base homeostasis and serotonin: A perspective on the use of carbon dioxide as human and rodent experimental model of panic. *Prog Neurobiol* 2015, 129: 58–78.
93. Khalsa SS, Adolphs R, Cameron OG, Critchley HD, Davenport PW, Feinstein JS, *et al.* Interoception and mental health: A roadmap. *Biol Psychiatry Cogn Neurosci Neuroimaging* 2018, 3: 501–513.
94. Clark DM. A cognitive approach to panic. *Behav Res Ther* 1986, 24: 461–470.
95. Domschke K, Stevens S, Pfleiderer B, Gerlach AL. Interoceptive sensitivity in anxiety and anxiety disorders: An overview and integration of neurobiological findings. *Clin Psychol Rev* 2010, 30: 1–11.
96. Gorman JM, Kent JM, Sullivan GM, Coplan JD. Neuroanatomical hypothesis of panic disorder, revised. *Am J Psychiatry* 2000, 157: 493–505.
97. Iversen SD. 5-HT and anxiety. *Neuropharmacology* 1984, 23: 1553–1560.
98. Kahn RS, Asnis GM, Wetzler S, van Praag HM. Neuroendocrine evidence for serotonin receptor hypersensitivity in panic disorder. *Psychopharmacology* 1988, 96: 360–364.
99. Deakin JF, Graeff FG. 5-HT and mechanisms of defence. *J Psychopharmacol* 1991, 5: 305–315.
100. Abell S. Serotonin-mediated anxiety: How to recognize and treat it. *Curr Psychiatry* 2021, 20: 37–40.
101. Coplan JD, Gorman JM, Klein DF. Serotonin related functions in panic-anxiety: A critical overview. *Neuropsychopharmacology* 1992, 6: 189–200.
102. Kahn RS, van Praag HM, Wetzler S, Asnis GM, Barr G. Serotonin and anxiety revisited. *Biol Psychiatry* 1988, 23: 189–208.
103. Maron E, Kuikka JT, Shlik J, Vasar V, Vanninen E, Tiitonen J. Reduced brain serotonin transporter binding in patients with panic disorder. *Psychiatry Res* 2004, 132: 173–181.
104. Neumeister A, Bain E, Nugent AC, Carson RE, Bonne O, Luckenbaugh DA, *et al.* Reduced serotonin type 1A receptor binding in panic disorder. *J Neurosci* 2004, 24: 589–591.
105. Šilhán P, Jelínková M, Walter U, Pavlov Praško J, Herzig R, Langová K, *et al.* Transcranial sonography of brainstem structures in panic disorder. *Psychiatry Res* 2015, 234: 137–143.
106. Zwanzger P, Rupprecht R. Selective GABAergic treatment for panic? Investigations in experimental panic induction and panic disorder. *J Psychiatry Neurosci* 2005, 30: 167–175.
107. Malizia AL, Cunningham VJ, Bell CJ, Liddle PF, Jones T, Nutt DJ. Decreased brain GABA(A)-benzodiazepine receptor binding in panic disorder: Preliminary results from a quantitative PET study. *Arch Gen Psychiatry* 1998, 55: 715–720.
108. Goddard AW, Mason GF, Almai A, Rothman DL, Behar KL, Petroff OA, *et al.* Reductions in occipital cortex GABA levels in panic disorder detected with 1h-magnetic resonance spectroscopy. *Arch Gen Psychiatry* 2001, 58: 556–561.
109. Sobanski T, Wagner G. Functional neuroanatomy in panic disorder: Status quo of the research. *World J Psychiatry* 2017, 7: 12–33.
110. Strawn JR, Levine A. Treatment response biomarkers in anxiety disorders: From neuroimaging to neuronally-derived extracellular vesicles and beyond. *Biomark Neuropsychiatry* 2020, 3: 100024.
111. Charney DS, Heninger GR, Breier A. Noradrenergic function in panic anxiety. Effects of yohimbine in healthy subjects and patients with agoraphobia and panic disorder. *Arch Gen Psychiatry* 1984, 41: 751–763.
112. Park HJ, Kim SK, Kang WS, Kim YJ, Cho AR, Park JK. Potential involvement of *NET* polymorphism in serotonin/norepinephrine reuptake inhibitor response in panic disorder. *Nord J Psychiatry* 2016, 70: 314–317.
113. Schunck T, Erb G, Mathis A, Gilles C, Namer IJ, Hode Y, *et al.* Functional magnetic resonance imaging characterization of CCK-4-induced panic attack and subsequent anticipatory anxiety. *Neuroimage* 2006, 31: 1197–1208.
114. Knott V, Mahoney C, Bradwejn J, Shlik J, Gunnarsson T. Effects of acute cholecystokinin infusion on hemispheric EEG asymmetry and coherence in healthy volunteers. *Prog Neuro Psychopharmacol Biol Psychiatry* 2003, 27: 179–184.
115. Zifra M. Panic disorder: A review of treatment options. *Ann Clin Psychiatry* 2021, 33: 124–133.
116. Klein DF. Delineation of two drug-responsive anxiety syndromes. *Psychopharmacologia* 1964, 5: 397–408.

117. Pollack MH, Allgulander C, Bandelow B, Cassano GB, Greist JH, Hollander E, *et al.* WCA recommendations for the long-term treatment of panic disorder. *CNS Spectr* 2003, 8: 17–30.
118. Batelaan NM, Van Balkom AJLM, Stein DJ. Evidence-based pharmacotherapy of panic disorder: An update. *Int J Neuropsychopharmacol* 2012, 15: 403–415.
119. Quagliato LA, Freire RC, Nardi AE. Risks and benefits of medications for panic disorder: A comparison of SSRIs and benzodiazepines. *Expert Opin Drug Saf* 2018, 17: 315–324.
120. Laux G (2021) Serotonin reuptake inhibitors: Citalopram, escitalopram, fluoxetine, fluvoxamine, paroxetine, and sertraline. *NeuroPsychopharmacotherapy*, Springer, Cham, pp 1–13.
121. Pollack MH, Lepola U, Koponen H, Simon NM, Worthington JJ, Emilien G, *et al.* A double-blind study of the efficacy of venlafaxine extended-release, paroxetine, and placebo in the treatment of panic disorder. *Depress Anxiety* 2007, 24: 1–14.
122. Ferguson JM, Khan A, Mangano R, Entsuah R, Tzanis E. Relapse prevention of panic disorder in adult outpatient responders to treatment with venlafaxine extended release. *J Clin Psychiatry* 2007, 68: 58–68.
123. Cameron OG, Huang GC, Nichols T, Koeppe RA, Minoshima S, Rose D, *et al.* Reduced gamma-aminobutyric acid(A)-benzodiazepine binding sites in insular cortex of individuals with panic disorder. *Arch Gen Psychiatry* 2007, 64: 793–800.
124. Möhler H. The GABA system in anxiety and depression and its therapeutic potential. *Neuropharmacology* 2012, 62: 42–53.
125. Bighelli I, Trespidi C, Castellazzi M, Cipriani A, Furukawa TA, Girlanda F, *et al.* Antidepressants and benzodiazepines for panic disorder in adults. *Cochrane Database Syst Rev* 2016, 9: CD011567.
126. Pecknold JC. Alprazolam in panic disorder and agoraphobia: Results from a multicenter trial. *Arch Gen Psychiatry* 1988, 45: 429–436.
127. Susman J, Klee B. The role of high-potency benzodiazepines in the treatment of panic disorder. *Prim Care Companion J Clin Psychiatry* 2005, 7: 5–11.
128. Kumar B, Gupta VP, Kumar V. A perspective on monoamine oxidase enzyme as drug target: Challenges and opportunities. *Curr Drug Targets* 2017, 18: 87–97.
129. Bruce SE, Vasile RG, Goisman RM, Salzman C, Spencer M, Machan JT, *et al.* Are benzodiazepines still the medication of choice for patients with panic disorder with or without agoraphobia? *Am J Psychiatry* 2003, 160: 1432–1438.
130. Schlicht KF, Mann K, Jungmann F, Kaes J, Post F, Münzel T, *et al.* A 48-year-old woman with panic attacks. *Lancet* 2014, 384: 280.
131. Hans E, Hiller W. A meta-analysis of nonrandomized effectiveness studies on outpatient cognitive behavioral therapy for adult anxiety disorders. *Clin Psychol Rev* 2013, 33: 954–964.
132. Norton PJ, Price EC. A meta-analytic review of adult cognitive-behavioral treatment outcome across the anxiety disorders. *J Nerv Ment Dis* 2007, 195: 521–531.
133. Watts SE, Turnell A, Kladnitski N, Newby JM, Andrews G. Treatment-as-usual (TAU) is anything but usual: A meta-analysis of CBT versus TAU for anxiety and depression. *J Affect Disord* 2015, 175: 152–167.
134. Kircher T, Arolt V, Jansen A, Pyka M, Reinhardt I, Kellermann T, *et al.* Effect of cognitive-behavioral therapy on neural correlates of fear conditioning in panic disorder. *Biol Psychiatry* 2013, 73: 93–101.
135. Katzman MA, Bleau P, Blier P, Chokka P, Kjernisted K, Van Ameringen M, *et al.* Canadian clinical practice guidelines for the management of anxiety, posttraumatic stress and obsessive-compulsive disorders. *BMC Psychiatry* 2014, 14: S1.
136. Pompili A, Furukawa TA, Efthimiou O, Imai H, Tajika A, Salanti G. Dismantling cognitive-behaviour therapy for panic disorder: A systematic review and component network meta-analysis. *Psychol Med* 2018, 48: 1945–1953.
137. Sánchez-Meca J, Rosa-Alcázar AI, Marín-Martínez F, Gómez-Conesa A. Psychological treatment of panic disorder with or without agoraphobia: A meta-analysis. *Clin Psychol Rev* 2010, 30: 37–50.
138. Boettcher H, Brake CA, Barlow DH. Origins and outlook of interoceptive exposure. *J Behav Ther Exp Psychiatry* 2016, 53: 41–51.
139. Griez E, van den Hout MA. Treatment of phobophobia by exposure to CO₂-induced anxiety symptoms. *J Nerv Ment Dis* 1983, 171: 506–508.
140. Griez E, van Den Hout MA. CO₂ inhalation in the treatment of panic attacks. *Behav Res Ther* 1986, 24: 145–150.
141. van den Hout M, Griez E. Some remarks on the nosology of anxiety states and panic disorders. *Acta Psychiatr Belg* 1983, 83: 33–42.
142. Van den Hout MA, Griez E. Panic symptoms after inhalation of carbon dioxide. *Br J Psychiatry* 1984, 144: 503–507.
143. Johnson PL, Federici LM, Shekhar A. Etiology, triggers and neurochemical circuits associated with unexpected, expected, and laboratory-induced panic attacks. *Neurosci Biobehav Rev* 2014, 46: 429–454.
144. Wenger S. Anesthesia and analgesia in rabbits and rodents. *J Exot Pet Med* 2012, 21: 7–16.
145. Spiaci A Jr, Vilela-Costa HH, Sant’Ana AB, Fernandes GG, Tercino Frias A, da Silva GSF, *et al.* Panic-like escape response elicited in mice by exposure to CO₂, but not hypoxia. *Prog Neuro Psychopharmacol Biol Psychiatry* 2018, 81: 178–186.
146. Blanchard DC, Griebel G, Blanchard RJ. Mouse defensive behaviors: Pharmacological and behavioral assays for anxiety and panic. *Neurosci Biobehav Rev* 2001, 25: 205–218.
147. Blanchard DC, Blanchard RJ. Ethoexperimental approaches to the biology of emotion. *Annu Rev Psychol* 1988, 39: 43–68.
148. Zangrossi H Jr, Graeff FG. Serotonin in anxiety and panic: Contributions of the elevated T-maze. *Neurosci Biobehav Rev* 2014, 46: 397–406.
149. Ziemann AE, Allen JE, Dahdaleh NS, Drebot II, Coryell MW, Wunsch AM, *et al.* The amygdala is a chemosensor that detects carbon dioxide and acidosis to elicit fear behavior. *Cell* 2009, 139: 1012–1021.
150. Smoller JW, Gallagher PJ, Duncan LE, McGrath LM, Haddad SA, Holmes AJ, *et al.* The human ortholog of acid-sensing ion channel gene ASIC1a is associated with panic disorder and amygdala structure and function. *Biol Psychiatry* 2014, 76: 902–910.
151. Johnson PL, Molosh A, Fitz SD, Truitt WA, Shekhar A. Orexin, stress, and anxiety/panic states. *Prog Brain Res* 2012, 198: 133–161.
152. Salvatore G, Bonaventure P, Shekhar A, Johnson PL, Lord B, Shireman BT, *et al.* Translational evaluation of novel selective orexin-1 receptor antagonist JNJ-61393215 in an experimental model for panic in rodents and humans. *Transl Psychiatry* 2020, 10: 308.
153. Blanchard RJ, Blanchard DC, Agullana R, Weiss SM. Twenty-two kHz alarm cries to presentation of a predator, by laboratory rats living in visible burrow systems. *Physiol Behav* 1991, 50: 967–972.
154. Beckett SG, Aspley S, Graham M, Marsden CA. Pharmacological manipulation of ultrasound induced defence behaviour in the rat. *Psychopharmacology* 1996, 127: 384–390.
155. Beckett SR, Duxon MS, Aspley S, Marsden CA. Central c-fos expression following 20kHz/ultrasound induced defence behaviour in the rat. *Brain Res Bull* 1997, 42: 421–426.
156. Klein S, Nicolas LB, Lopez-Lopez C, Jacobson LH, McArthur SG, Grundschober C, *et al.* Examining face and construct

- validity of a noninvasive model of panic disorder in Lister-hooded rats. *Psychopharmacology* 2010, 211: 197–208.
157. Schenberg LC, Bittencourt AS, Sudré EC, Vargas LC. Modeling panic attacks. *Neurosci Biobehav Rev* 2001, 25: 647–659.
 158. Nicolas LB, Klein S, Prinssen EP. Defensive-like behaviors induced by ultrasound: Further pharmacological characterization in Lister-hooded rats. *Psychopharmacology* 2007, 194: 243–252.
 159. Blanchard DC, Griebel G, Blanchard RJ. The mouse defense test battery: Pharmacological and behavioral assays for anxiety and panic. *Eur J Pharmacol* 2003, 463: 97–116.
 160. Griebel G, Blanchard DC, Agnes RS, Blanchard RJ. Differential modulation of antipredator defensive behavior in Swiss-Webster mice following acute or chronic administration of imipramine and fluoxetine. *Psychopharmacology* 1995, 120: 57–66.
 161. Griebel G, Blanchard DC, Jung A, Blanchard RJ. A model of 'antipredator' defense in Swiss-Webster mice: Effects of benzodiazepine receptor ligands with different intrinsic activities. *Behav Pharmacol* 1995, 6: 732–745.
 162. Pobbe RL, Zangrossi H Jr, Blanchard DC, Blanchard RJ. Involvement of dorsal raphe nucleus and dorsal periaqueductal gray 5-HT receptors in the modulation of mouse defensive behaviors. *Eur Neuropsychopharmacol* 2011, 21: 306–315.
 163. Blanchard DC, Griebel G, Blanchard RJ. Conditioning and residual emotionality effects of predator stimuli: Some reflections on stress and emotion. *Prog Neuropsychopharmacol Biol Psychiatry* 2003, 27: 1177–1185.
 164. Griebel G, Blanchard DC, Jung A, Lee JC, Masuda CK, Blanchard RJ. Further evidence that the mouse defense test battery is useful for screening anxiolytic and panicolytic drugs: Effects of acute and chronic treatment with alprazolam. *Neuropharmacology* 1995, 34: 1625–1633.
 165. Griebel G, Beeské S (2011) The mouse defense test battery: A model measuring different facets of anxiety-related behaviors. In: Gould TD (ed) *Mood and Anxiety Related Phenotypes in Mice*, Humana Press, Totowa, pp 97–106.
 166. Pellow S, Chopin P, File SE, Briley M. Validation of open: Closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J Neurosci Methods* 1985, 14: 149–167.
 167. Zangrossi H Jr, Graeff FG. Behavioral validation of the elevated T-maze, a new animal model of anxiety. *Brain Res Bull* 1997, 44: 1–5.
 168. Pinheiro SN, Del-Ben CM, Zangrossi H Jr, Graeff FG. Anxiolytic and panicolytic effects of escitalopram in the elevated T-maze. *J Psychopharmacol* 2008, 22: 132–137.
 169. Teixeira RC, Zangrossi H, Graeff FG. Behavioral effects of acute and chronic imipramine in the elevated T-maze model of anxiety. *Pharmacol Biochem Behav* 2000, 65: 571–576.
 170. Dos Anjos Rosário B, de Fátima Santana de Nazaré M, Lemes JA, de Andrade JS, da Silva RB, Pereira CDS, *et al.* Repeated crack cocaine administration alters panic-related responses and delta FosB immunoreactivity in panic-modulating brain regions. *Exp Brain Res* 2021, 239: 1179–1191.
 171. Jardim MC, Nogueira RL, Graeff FG, Nunes-de-Souza RL. Evaluation of the elevated T-maze as an animal model of anxiety in the mouse. *Brain Res Bull* 1999, 48: 407–411.
 172. Gorman JM, Liebowitz MR, Fyer AJ, Stein J. A neuroanatomical hypothesis for panic disorder. *Am J Psychiatry* 1989, 146: 148–161.
 173. Lai CH. Fear network model in panic disorder: The past and the future. *Psychiatry Invest* 2019, 16: 16–26.
 174. Maren S. An acid-sensing channel sows fear and panic. *Cell* 2009, 139: 867–869.
 175. Wemmie JA. Neurobiology of panic and pH chemosensation in the brain. *Dialogues Clin Neurosci* 2011, 13: 475–483.
 176. Taugher RJ, Dlouhy BJ, Kreple CJ, Ghobbeh A, Conlon MM, Wang Y, *et al.* The amygdala differentially regulates defensive behaviors evoked by CO₂. *Behav Brain Res* 2020, 377: 112236.
 177. Xu L. Leptin action in the midbrain: From reward to stress. *J Chem Neuroanat* 2014, 61(62): 256–265.
 178. Vagnoni E, Lourenco SF, Longo MR. Threat modulates perception of looming visual stimuli. *Curr Biol* 2012, 22: R826–R827.
 179. Gale SD, Murphy GJ. Distinct representation and distribution of visual information by specific cell types in mouse superficial superior colliculus. *J Neurosci* 2014, 34: 13458–13471.
 180. Yilmaz M, Meister M. Rapid innate defensive responses of mice to looming visual stimuli. *Curr Biol* 2013, 23: 2011–2015.
 181. de Vries SE, Clandinin TR. Loom-sensitive neurons link computation to action in the *Drosophila* visual system. *Curr Biol* 2012, 22: 353–362.
 182. Dunn TW, Gebhardt C, Naumann EA, Riegler C, Ahrens MB, Engert F, *et al.* Neural circuits underlying visually evoked escapes in larval zebrafish. *Neuron* 2016, 89: 613–628.
 183. Temizer I, Donovan JC, Baier H, Semmelhack JL. A visual pathway for looming-evoked escape in larval zebrafish. *Curr Biol* 2015, 25: 1823–1834.
 184. Huang L, Yuan T, Tan M, Xi Y, Hu Y, Tao Q, *et al.* A retinoraphe projection regulates serotonergic activity and looming-evoked defensive behaviour. *Nat Commun* 2017, 8: 14908.
 185. Liu YJ, Wang Q, Li B. Neuronal responses to looming objects in the superior colliculus of the cat. *Brain Behav Evol* 2011, 77: 193–205.
 186. Evans DA, Stempel AV, Vale R, Ruehle S, Lefler Y, Branco T. A synaptic threshold mechanism for computing escape decisions. *Nature* 2018, 558: 590–594.
 187. Shang C, Liu Z, Chen Z, Shi Y, Wang Q, Liu S, *et al.* BRAIN CIRCUITS. A parvalbumin-positive excitatory visual pathway to trigger fear responses in mice. *Science* 2015, 348: 1472–1477.
 188. Wei P, Liu N, Zhang Z, Liu X, Tang Y, He X, *et al.* Processing of visually evoked innate fear by a non-canonical thalamic pathway. *Nat Commun* 2015, 6: 6756.
 189. Shang C, Chen Z, Liu A, Li Y, Zhang J, Qu B, *et al.* Divergent midbrain circuits orchestrate escape and freezing responses to looming stimuli in mice. *Nat Commun* 2018, 9: 1232.
 190. Xie Z, Wang M, Liu Z, Shang C, Zhang C, Sun L, *et al.* Transcriptomic encoding of sensorimotor transformation in the midbrain. *Elife* 2021, 10: e69825.
 191. Liang F, Xiong XR, Zingg B, Ji XY, Zhang LI, Tao HW. Sensory cortical control of a visually induced arrest behavior via corticotectal projections. *Neuron* 2015, 86: 755–767.
 192. Zhao X, Liu M, Cang J. Visual cortex modulates the magnitude but not the selectivity of looming-evoked responses in the superior *Colliculus* of awake mice. *Neuron* 2014, 84: 202–213.
 193. Carrive P. The periaqueductal gray and defensive behavior: Functional representation and neuronal organization. *Behav Brain Res* 1993, 58: 27–47.
 194. Gross CT, Canteras NS. The many paths to fear. *Nat Rev Neurosci* 2012, 13: 651–658.
 195. Deng H, Xiao X, Wang Z. Periaqueductal gray neuronal activities underlie different aspects of defensive behaviors. *J Neurosci* 2016, 36: 7580–7588.
 196. Esteban Masferrer M, Silva BA, Nomoto K, Lima SQ, Gross CT. Differential encoding of predator fear in the ventromedial hypothalamus and periaqueductal grey. *J Neurosci* 2020, 40: 9283–9292.
 197. Marchand JE, Hagino N. Afferents to the periaqueductal gray in the rat. A horseradish peroxidase study. *Neuroscience* 1983, 9: 95–106.
 198. Tovote P, Esposito MS, Botta P, Chaudun F, Fadok JP, Markovic M, *et al.* Midbrain circuits for defensive behaviour. *Nature* 2016, 534: 206–212.

199. Silva BA, Mattucci C, Krzykowski P, Murana E, Illarionova A, Grinevich V, *et al.* Independent hypothalamic circuits for social and predator fear. *Nat Neurosci* 2013, 16: 1731–1733.
200. Hahn JD, Sporns O, Watts AG, Swanson LW. Macroscale intrinsic network architecture of the hypothalamus. *Proc Natl Acad Sci U S A* 2019, 116: 8018–8027.
201. Li Y, Zeng J, Zhang J, Yue C, Zhong W, Liu Z, *et al.* Hypothalamic circuits for predation and evasion. *Neuron* 2018, 97: 911–924.e5.
202. Xiong XR, Liang F, Zingg B, Ji XY, Ibrahim LA, Tao HW, *et al.* Auditory cortex controls sound-driven innate defense behaviour through corticofugal projections to inferior colliculus. *Nat Commun* 2015, 6: 7224.
203. Chou XL, Wang X, Zhang ZG, Shen L, Zingg B, Huang J, *et al.* Inhibitory gain modulation of defense behaviors by zona incerta. *Nat Commun* 2018, 9: 1151.
204. Ferreira-Pinto MJ, Ruder L, Capelli P, Arber S. Connecting circuits for supraspinal control of locomotion. *Neuron* 2018, 100: 361–374.
205. Capelli P, Pivetta C, Soledad Esposito M, Arber S. Locomotor speed control circuits in the caudal brainstem. *Nature* 2017, 551: 373–377.
206. Kim EJ, Horovitz O, Pellman BA, Tan LM, Li Q, Richter-Levin G, *et al.* Dorsal periaqueductal gray-amygdala pathway conveys both innate and learned fear responses in rats. *Proc Natl Acad Sci U S A* 2013, 110: 14795–14800.
207. Schmitt P, Sandner G, Karli P. Escape and approach induced by brain stimulation: A parametric analysis. *Behav Brain Res* 1981, 2: 49–79.
208. Jenck F, Moreau JL, Martin JR. Dorsal periaqueductal gray-induced aversion as a simulation of panic anxiety: Elements of face and predictive validity. *Psychiatry Res* 1995, 57: 181–191.
209. Del-Ben CM, Graeff FG. Panic disorder: Is the PAG involved? *Neural Plast* 2009, 2009: 108135.
210. Kiser RS, Lebovitz RM. Monoaminergic mechanisms in aversive brain stimulation. *Physiol Behav* 1975, 15: 47–53.
211. Schenberg LC, Graeff FG. Role of the periaqueductal gray substance in the antianxiety action of benzodiazepines. *Pharmacol Biochem Behav* 1978, 9: 287–295.
212. Wise CD, Berger BD, Stein L. Benzodiazepines: Anxiety-reducing activity by reduction of serotonin turnover in the brain. *Science* 1972, 177: 180–183.
213. Kiser RS, German DC, Lebovitz RM. Serotonergic reduction of dorsal central gray area stimulation-produced aversion. *Pharmacol Biochem Behav* 1978, 9: 27–31.
214. Sanford Kiser R, Brown CA, Sanghera MK, German DC. Dorsal raphe nucleus stimulation reduces centrally-elicited fearful behavior. *Brain Res* 1980, 191: 265–272.
215. Canteras NS, Ribeiro-Barbosa ER, Comoli E. Tracing from the dorsal premammillary nucleus prosencephalic systems involved in the organization of innate fear responses. *Neurosci Biobehav Rev* 2001, 25: 661–668.
216. Graeff FG (1991) Neurotransmitters in the dorsal periaqueductal grey and animal models of panic anxiety. In: Briley M, File SE (eds) *New Concepts in Anxiety*, Macmillan Education UK, London, pp 288–312.
217. Bortoli VC, Nogueira RL, Zangrossi H Jr. Effects of fluoxetine and buspirone on the panicolytic-like response induced by the activation of 5-HT_{1A} and 5-HT_{2A} receptors in the rat dorsal periaqueductal gray. *Psychopharmacology* 2006, 183: 422–428.
218. Hess WR, Akert K. Experimental data on role of hypothalamus in mechanism of emotional behavior. *AMA Arch Neurol Psychiatry* 1955, 73: 127–129.
219. Hess WR, Brügger M. Das subkortikale zentrum der affektiven abwehrreaktion. *Helv Physiol Pharmacol Acta* 1943, 1: 33–52.
220. Fuchs SA, Edinger HM, Siegel A. The organization of the hypothalamic pathways mediating affective defense behavior in the cat. *Brain Res* 1985, 330: 77–92.
221. Yardley CP, Hilton SM. The hypothalamic and brainstem areas from which the cardiovascular and behavioural components of the defence reaction are elicited in the rat. *J Auton Nerv Syst* 1986, 15: 227–244.
222. Canteras NS, Chiavegatto S, Ribeiro do Valle LE, Swanson LW. Severe reduction of rat defensive behavior to a predator by discrete hypothalamic chemical lesions. *Brain Res Bull* 1997, 44: 297–305.
223. Choi GB, Dong HW, Murphy AJ, Valenzuela DM, Yancopoulos GD, Swanson LW, *et al.* *Lhx6* delineates a pathway mediating innate reproductive behaviors from the amygdala to the hypothalamus. *Neuron* 2005, 46: 647–660.
224. Bergan JF, Ben-Shaul Y, Dulac C. Sex-specific processing of social cues in the medial amygdala. *Elife* 2014, 3: e02743.
225. Lin D, Boyle MP, Dollar P, Lee H, Lein ES, Perona P, *et al.* Functional identification of an aggression locus in the mouse hypothalamus. *Nature* 2011, 470: 221–226.
226. Wang L, Chen IZ, Lin D. Collateral pathways from the ventromedial hypothalamus mediate defensive behaviors. *Neuron* 2015, 85: 1344–1358.
227. Silva BA, Mattucci C, Krzykowski P, Cuzzo R, Carbonari L, Gross CT. The ventromedial hypothalamus mediates predator fear memory. *Eur J Neurosci* 2016, 43: 1431–1439.
228. Kunwar PS, Zelikowsky M, Remedios R, Cai H, Yilmaz M, Meister M, *et al.* Ventromedial hypothalamic neurons control a defensive emotion state. *Elife* 2015, 4: e06633.
229. Lindberg D, Chen P, Li C. Conditional viral tracing reveals that steroidogenic factor 1-positive neurons of the dorsomedial subdivision of the ventromedial hypothalamus project to autonomic centers of the hypothalamus and hindbrain. *J Comp Neurol* 2013, 521: 3167–3190.
230. Staples LG, Hunt GE, Cornish JL, McGregor IS. Neural activation during cat odor-induced conditioned fear and ‘trial 2’ fear in rats. *Neurosci Biobehav Rev* 2005, 29: 1265–1277.
231. Canteras NS, Kroon JA, Do-Monte FH, Pavesi E, Carobrez AP. Sensing danger through the olfactory system: The role of the hypothalamic dorsal premammillary nucleus. *Neurosci Biobehav Rev* 2008, 32: 1228–1235.
232. Cezario AF, Ribeiro-Barbosa ER, Baldo MV, Canteras NS. Hypothalamic sites responding to predator threats—the role of the dorsal premammillary nucleus in unconditioned and conditioned antipredatory defensive behavior. *Eur J Neurosci* 2008, 28: 1003–1015.
233. Wang W, Schuette PJ, Jun N, Tobias BC, Cuccovia V, Reis FM, Ji S, *et al.* Coordination of escape and spatial navigation circuits orchestrates versatile flight from threats. *Neuron* 2021, 109: 1848–1860.e8.
234. Paschoalin-Maurin T, Dos Anjos-Garcia T, Falconi-Sobrinho LL, de Freitas RL, Coimbra JPC, Laure CJ, *et al.* The rodent-versus-wild snake paradigm as a model for studying anxiety- and panic-like behaviors: Face, construct and predictive validities. *Neuroscience* 2018, 369: 336–349.
235. Lynn R. National rates of economic growth, anxiety and suicide. *Nature* 1969, 222: 494.
236. Markgraf CG, Winters RW, Liskowsky DR, McCabe PM, Green EJ, Schneiderman N. Hypothalamic, midbrain and bulbar areas involved in the defense reaction in rabbits. *Physiol Behav* 1991, 49: 493–500.
237. Johnson PL, Shekhar A. An animal model of panic vulnerability with chronic disinhibition of the dorsomedial/perifornical hypothalamus. *Physiol Behav* 2012, 107: 686–698.
238. Shekhar A, Keim SR, Simon JR, McBride WJ. Dorsomedial hypothalamic GABA dysfunction produces physiological

- arousal following sodium lactate infusions. *Pharmacol Biochem Behav* 1996, 55: 249–256.
239. Shekhar A, Keim SR. The circumventricular organs form a potential neural pathway for lactate sensitivity: Implications for panic disorder. *J Neurosci* 1997, 17: 9726–9735.
 240. Shekhar A, Johnson PL, Fitz SD, Nakazato A, Chaki S, Steckler T, *et al.* A selective, non-peptide CRF receptor 1 antagonist prevents sodium lactate-induced acute panic-like responses. *Int J Neuropsychopharmacol* 2011, 14: 355–365.
 241. Soltis RP, DiMicco JA. Interaction of hypothalamic GABAA and excitatory amino acid receptors controlling heart rate in rats. *Am J Physiol* 1991, 261: R427–R433.
 242. Johnson PL, Shekhar A. Panic-prone state induced in rats with GABA dysfunction in the dorsomedial hypothalamus is mediated by NMDA receptors. *J Neurosci* 2006, 26: 7093–7104.
 243. Shekhar A, Keim SR. LY354740, a potent group II metabotropic glutamate receptor agonist prevents lactate-induced panic-like response in panic-prone rats. *Neuropharmacology* 2000, 39: 1139–1146.
 244. Schoepp DD, Wright RA, Levine LR, Gaydos B, Potter WZ. LY354740, an mGlu2/3 receptor agonist as a novel approach to treat anxiety/stress. *Stress* 2003, 6: 189–197.
 245. de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, *et al.* The hypocretins: Hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci U S A* 1998, 95: 322–327.
 246. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, *et al.* Orexins and orexin receptors: A family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 1998, 92: 573–585.
 247. Thannickal TC, Lai YY, Siegel JM. Hypocretin (orexin) cell loss in Parkinson's disease. *Brain* 2007, 130: 1586–1595.
 248. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, *et al.* Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* 1998, 18: 9996–10015.
 249. Han Y, Yuan K, Zheng Y, Lu L. Orexin receptor antagonists as emerging treatments for psychiatric disorders. *Neurosci Bull* 2020, 36: 432–448.
 250. Johnson PL, Truitt W, Fitz SD, Minick PE, Dietrich A, Sanghani S, *et al.* A key role for orexin in panic anxiety. *Nat Med* 2010, 16: 111–115.
 251. Bromberg-Martin ES, Matsumoto M, Hikosaka O. Dopamine in motivational control: Rewarding, aversive, and alerting. *Neuron* 2010, 68: 815–834.
 252. Schultz W. Dopamine reward prediction-error signalling: A two-component response. *Nat Rev Neurosci* 2016, 17: 183–195.
 253. Ciochi S, Herry C, Grenier F, Wolff SBE, Letzkus JJ, Vlachos I, *et al.* Encoding of conditioned fear in central amygdala inhibitory circuits. *Nature* 2010, 468: 277–282.
 254. Haubensak W, Kunwar PS, Cai H, Ciochi S, Wall NR, Ponnusamy R, *et al.* Genetic dissection of an amygdala microcircuit that gates conditioned fear. *Nature* 2010, 468: 270–276.
 255. Moriya S, Yamashita A, Kawashima S, Nishi R, Yamanaka A, Kuwaki T. Acute aversive stimuli rapidly increase the activity of ventral tegmental area dopamine neurons in awake mice. *Neuroscience* 2018, 386: 16–23.
 256. Nieh EH, Vander Weele CM, Matthews GA, Presbrey KN, Wichmann R, Leppla CA, *et al.* Inhibitory input from the lateral hypothalamus to the ventral tegmental area disinhibits dopamine neurons and promotes behavioral activation. *Neuron* 2016, 90: 1286–1298.
 257. Zhou Z, Liu X, Chen S, Zhang Z, Liu Y, Montardy Q, *et al.* A VTA GABAergic neural circuit mediates visually evoked innate defensive responses. *Neuron* 2019, 103: 473–488.e6.
 258. Barbano MF, Wang HL, Zhang S, Miranda-Barrientos J, Estrin DJ, Figueroa-González A, *et al.* VTA glutamatergic neurons mediate innate defensive behaviors. *Neuron* 2020, 107: 368–382.e8.
 259. Shik ML, Severin FV, Orlovskii GN. Control of walking and running by means of electric stimulation of the midbrain. *Biofizika* 1966, 11: 659–666.
 260. Ryczko D, Dubuc R. The multifunctional mesencephalic locomotor region. *Curr Pharm Des* 2013, 19: 4448–4470.
 261. Roseberry TK, Lee AM, Lalive AL, Wilbrecht L, Bonci A, Kreitzer AC. Cell-type-specific control of brainstem locomotor circuits by basal Ganglia. *Cell* 2016, 164: 526–537.
 262. Lee AM, Hoy JL, Bonci A, Wilbrecht L, Stryker MP, Niell CM. Identification of a brainstem circuit regulating visual cortical state in parallel with locomotion. *Neuron* 2014, 83: 455–466.
 263. Giber K, Diana MA, Plattner V, Dugué GP, Bokor H, Rousseau CV, *et al.* A subcortical inhibitory signal for behavioral arrest in the thalamus. *Nat Neurosci* 2015, 18: 562–568.
 264. Halladay LR, Blair HT. Distinct ensembles of medial prefrontal cortex neurons are activated by threatening stimuli that elicit excitation vs. inhibition of movement. *J Neurophysiol* 2015, 114: 793–807.
 265. Vander Weele CM, Siciliano CA, Matthews GA, Namburi P, Izadmehr EM, Espinel IC, *et al.* Dopamine enhances signal-to-noise ratio in cortical-brainstem encoding of aversive stimuli. *Nature* 2018, 563: 397–401.
 266. Leonardo de Freitas R, Salgado-Rohner CJ, Biagioni AF, Medeiros P, Hallak JEC, Alexandre S, Crippa J, *et al.* NMDA and AMPA/kainate glutamatergic receptors in the prelimbic medial prefrontal cortex modulate the elaborated defensive behavior and innate fear-induced antinociception elicited by GABAA receptor blockade in the medial hypothalamus. *Cereb Cortex* 2014, 24: 1518–1528.
 267. Eichenbaum H. *Hippocampus*: Mapping or memory? *Curr Biol* 2000, 10: R785–R787.
 268. Wang ME, Fraize NP, Yin L, Yuan RK, Petsagourakis D, Wann EG, *et al.* Differential roles of the dorsal and ventral hippocampus in predator odor contextual fear conditioning. *Hippocampus* 2013, 23: 451–466.
 269. Berridge CW, Waterhouse BD. The locus coeruleus-noradrenergic system: Modulation of behavioral state and state-dependent cognitive processes. *Brain Res Rev* 2003, 42: 33–84.
 270. Carter ME, Yizhar O, Chikahisa S, Nguyen H, Adamantidis A, Nishino S, *et al.* Tuning arousal with optogenetic modulation of locus coeruleus neurons. *Nat Neurosci* 2010, 13: 1526–1533.
 271. Hickey L, Li Y, Fyson SJ, Watson TC, Perrins R, Hewinson J, *et al.* Optoactivation of locus ceruleus neurons evokes bidirectional changes in thermal nociception in rats. *J Neurosci* 2014, 34: 4148–4160.
 272. Li L, Feng X, Zhou Z, Zhang H, Shi Q, Lei Z, *et al.* Stress accelerates defensive responses to looming in mice and involves a locus coeruleus-superior Colliculus projection. *Curr Biol* 2018, 28: 859–871.e5.
 273. Tsuruoka M, Tamaki J, Maeda M, Hayashi B, Inoue T. The nucleus locus coeruleus/subcoeruleus contributes to antinociception during freezing behavior following the air-puff startle in rats. *Brain Res* 2011, 1393: 52–61.
 274. Duval ER, Javanbakht A, Liberzon I. Neural circuits in anxiety and stress disorders: A focused review. *Ther Clin Risk Manag* 2015, 11: 115–126.

275. Amano K, Tanikawa T, Iseki H, Kawabatake H, Notani M, Kawamura H, *et al.* Single neuron analysis of the human mid-brain tegmentum. Rostral mesencephalic reticulotomy for pain relief. *Appl Neurophysiol* 1978, 41: 66–78.
276. Wilent WB, Oh MY, Buetefisch CM, Bailes JE, Cantella D, Angle C, *et al.* Induction of panic attack by stimulation of the ventromedial hypothalamus. *J Neurosurg* 2010, 112: 1295–1298.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.